

Appendix H

Selected DEQ Tittabawassee River Sediment Study Results and Maps

-- Note --

This document is an abbreviated version of
the full 163-page report entitled:

Baseline Chemical Characterization of Saginaw Bay Watershed Sediments
The full report can be accessed from this DEQ web site:

<http://www.deq.state.mi.us/documents/deq-rrd-dioxin-FinalReport.pdf>

BASELINE CHEMICAL CHARACTERIZATION OF SAGINAW BAY WATERSHED SEDIMENTS

A Report to the Office of the Great Lakes
Michigan Department of Environmental Quality



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Baseline Chemical Characterization of Saginaw Bay Watershed Sediments

Introduction

General: Persistent bioaccumulative compounds have historically been released from industrial entities in the Saginaw Bay watershed. Dioxins and furans, polychlorinated and polybrominated biphenyl compounds (PCBs and PBBs), pesticides, herbicides, and other persistent bioaccumulative compounds have been identified as significant pollutants in the Saginaw River and the Saginaw Bay (U.S.EPA, 1995).

In September of 2000, the Waste Management Division of the Michigan Department of Environmental Quality (MDEQ) was awarded a grant of \$88,775.00 from the Michigan Great Lakes Protection Fund to conduct a study entitled: "Baseline Chemical Characterization of Saginaw Bay Watershed Sediments (Baseline Study)." The Baseline Study area is shown below as Figure 1. A copy of the approved grant proposal and budget is attached as Appendix 1 of this Report.

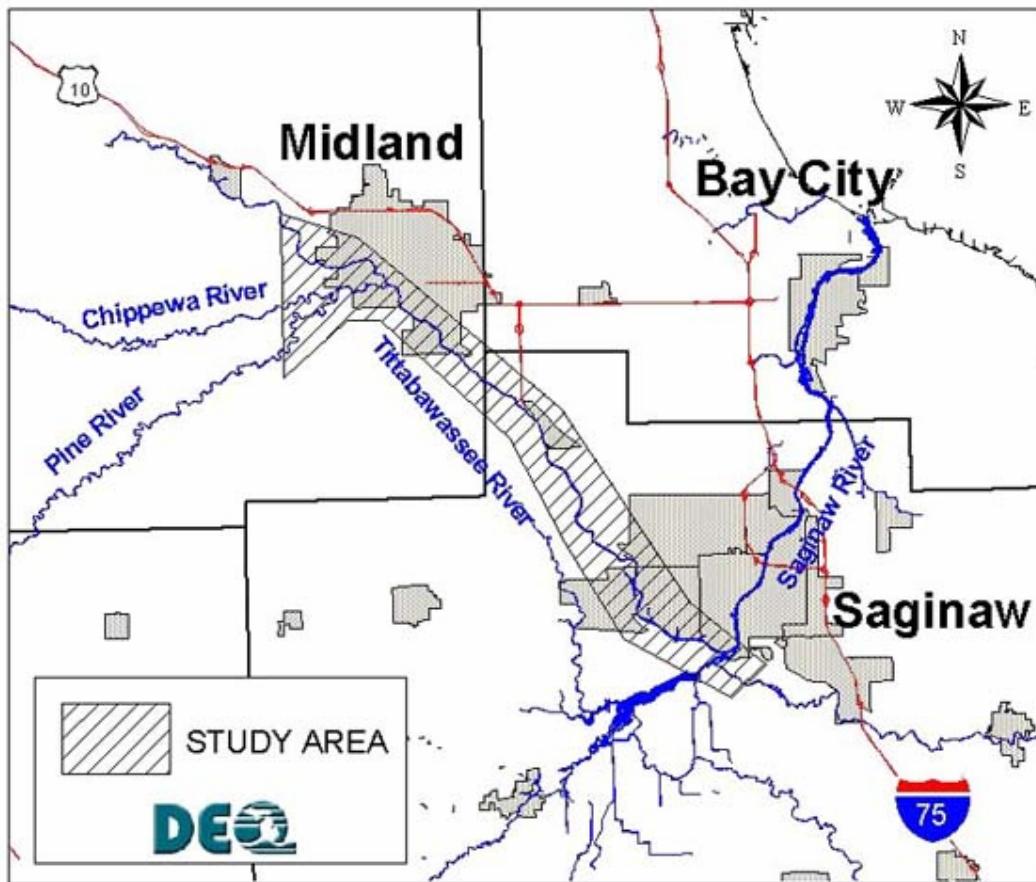


Figure 1

The focus of the Baseline Study is to characterize the sediments in the identified study area beginning upstream of Midland Michigan on the Pine, Chippewa, and Tittabawassee Rivers, and continuing downstream on the Tittabawassee to its confluence with the Saginaw River. The data and sampling locations from this study establish a year 2001 baseline level of contamination of watershed sediments that provides a benchmark against which future improvements in sediment and water quality can be measured. The analytical data from this project is also intended to serve as a screening level evaluation of sediment quality in the Tittabawassee River; to determine if contaminants are present at levels of environmental concern; and to determine if Tittabawassee River sediments are a potential source of ongoing releases to Lake Huron. Finally, analytical results from the study can be used to form the basis for a request for corrective action, if necessary, from regulated facilities within the watershed.

The Baseline Study was developed and conducted in a phased manner that consisted of a review of existing literature and data; the development of a sampling and analysis plan; the collection of samples; the analysis of the data; and the preparation of this Report. The Michigan State University Aquatic Toxicology Laboratory (MSU ATL), under the direction of Dr. John Giesy, assisted in the completion of this project by conducting a detailed literature review, assisting with the development of the study sampling and analysis plan (SAP), and conducting the dioxin and furan related analyses on the samples.

Literature and Data Review

The MSU-ATL conducted a literature search to identify appropriate literature to support the Baseline Study. This included all general literature on polychlorinated diaromatic hydrocarbons and specifically all of the literature, either in the open or gray literature and reports on the Saginaw River drainage system. The literature is compiled into an electronic data base (Reference Manager^Æ). A hard copy of the literature search is attached as Appendix 2 of this study.

The literature review indicates that significant sediment investigation work has been downstream of the study area in the Saginaw River and in the Saginaw Bay. In particular, the United States Environmental Protection Agency (U.S. EPA) has identified the Saginaw River and Saginaw Bay as an Area of Concern (U.S. EPA, 1995) as part of the Assessment and Remediation of Contaminated Sediments Program (ARCS Program). The ARCS Program resulted in the development of detailed sediment quality information on the lower Saginaw River and Saginaw Bay. Grab and core sediment samples from the ARCS study were analyzed for polyaromatic hydrocarbons, PCBs, chlorinated pesticides, polychlorinated dibenzo-p-

dioxins, polychlorinated dibenzofurans, and metals. General conclusions regarding the presence and risk of PCBs and certain metals in the sediments of the Area of Concern are presented in this study. In addition, to the ARCS Program, the Army Corps of Engineers (1999) has also conducted fairly extensive sampling and analysis of sediments in the navigable channel of the Saginaw associated with dredging projects.

Much less data is available on sediment quality in the Tittabawassee, Pine, and Chippewa Rivers upstream of the confluence of the Tittabawassee and the Saginaw Rivers. The Michigan Department of Community Health has issued fish advisories based on elevated levels of dioxins and furans and PCBs found in fish tissues in the Tittabawassee and Saginaw Rivers downstream of Midland. Fish advisories for PBBs and DDT have been issued for all species of fish on the Pine River downstream of the St. Louis impoundment.

Amendola and Barna (1986) reported of dioxin concentrations at up to 16 parts per billion (OCDD) in Tittabawassee River sediments in 1984. Dioxins were not detected in sediments upstream of The Dow Chemical Company facility in Midland, Michigan. Two studies have analyzed Tittabawassee River sediments for PCBs (MDNR, 1971, 1988). The Michigan Department of Natural Resources summaries of this work indicate that the sediment data from these studies is relatively sparse.

The MDEQ Surface Water Quality Division conducted an extensive review of existing information on the Tittabawassee River in 1993 during the evaluation of a potential Natural Resources Damage Assessment claim against The Dow Chemical Company (RCG/Hagler, Baily Inc., 1993). That review identified data gaps in water quality and sediment quality with respect to dioxins and furans and PCBs.

The MDEQ conducted limited sediment sampling of the Tittabawassee and Chippewa Rivers in 1996 for a broad range of organic compounds, including dioxins and furans, and metals (MDEQ, 1996). This sampling event was concentrated adjacent to The Dow Chemical Company facility and immediately upstream of the facility.

Based on a review of existing data, it was clear that no comprehensive sediment characterization program has been conducted on the Tittabawassee River and that there is no program which routinely analyzes sediments to track changes in environmental quality of this portion of the Saginaw Bay watershed. It was also apparent that dioxins and furans were a significant issue in this watershed.

Methodology

Sampling and Analysis Plan: A SAP was developed for the collection and analysis of the samples. A copy of the SAP is attached as Appendix 3 to this Report. Unless otherwise stated, analyses were conducted in accordance with "Test Methods for Evaluating Solid Waste, Physical/Chemical Methods," EPA Publication SW-846, Third Edition, November 1986, and its updates I (July 1992), II (September 1994), IIA (August 1993), IIB (January 1995), III (December 1996), and IIIA (April 1998) (SW-846)

Study Design: The goals of this project were to (1) obtain a baseline chemical characterization of Tittabawassee River sediments and floodplain soils; and (2) to choose and document sampling locations so that the study might be replicated at a future date. These goals, in concert with financial and logistical realities, resulted in the sampling and analysis approach utilized in this project.

The parameters of interest in this study consisted of heavy metals, volatile and semivolatile organic compounds, pesticides, PCBs, dioxins, and furans. These parameters were chosen by the principal investigators as being the best indicators of the overall environmental health of the river system. A complete list of individual parameters is included in Table 1. In addition, analyses of physical parameters [Total Organic Carbon (TOC) and Total Organic Matter] were performed on all samples.

In addition to conventional dioxin and furan instrumental analysis (SW-846 Method 8290), each of the transect, composite, floodplain soils, and selected individual reach samples described below were analyzed using H4IIE-luc bioassay to determine relative dioxin-like activity. A subset of these samples were further analyzed in order to perform mass balance calculations to determine if other compounds are contributing significant dioxin like activity to Tittabawassee River sediments and soils.

The persistent bioaccumulative compounds of concern in this study are most frequently associated with fine particulate matter in the environment. Due to this association, an attempt was made to preferentially sample areas of fine particulate deposition. Preferred sampling locations included zones immediately downstream of large snags, bridge pilings, or other man-made structures, the inside bend of river meanders and other areas where fine particulates were observed to accumulate. Field personnel frequently tested possible sampling locations by poling with a boat oar to determine if there was suitable sediment accumulation. Figure 2 illustrates typical river sampling locations.

The location of each sample collected during this study was memorialized using hand-held Global Positioning System technology. Location coordinates in decimal degrees latitude and longitude for each sampling location in this study are identified in Table 2.



Figure 2. Typical River Sample Locations

Transect Sampling Locations: Sampling locations were chosen to provide geographic coverage of the study area and to preferentially sample depositional areas where the parameters of concern were most likely to be found. Nine locations (hereinafter referred to as transects) were chosen in order to provide coverage along the approximately 22 miles of river comprising the study area. Transect locations were tentatively identified by reviewing maps of the study area and locations finalized by the identification of depositional zones in the field. Transect locations are shown in Figure 3.

Two types of sampling were typically performed at each transect location. A sediment core sample was obtained by driving a four-inch diameter acetate or polyvinyl chloride (PVC) tube into the river bottom to a depth of one and a half to two feet (0.45 to 0.6 meter) at one discrete location per transect. Stones larger than $\frac{1}{2}$ inch and leaves and twigs were removed from the sample before compositing in a stainless steel bowl. Aliquots of the composite sample were taken for analysis for each of the study parameters. Figure 4 illustrates a typical core sample.



Figure 4. Typical Core sample

A composite sample of the surficial sediments was obtained by using a petite Ponar dredge to collect a sample of the upper one to two inches of sediment at three to five locations per transect along a line between each bank of the river. The number of locations per transect was a field decision based on stream width, particle size of the sediment, and practical considerations in obtaining a sample. The individual dredge samples were composited by mixing with a stainless steel scoop in a

stainless steel bowl. Stones larger than $\frac{1}{8}$ inch and leaves and twigs were removed from the sample before compositing in a stainless steel bowl. Figure 5 illustrates a typical upper composite sample. Aliquots of the composite sample were taken for analysis for each of the study parameters.



Figure 5. Typical Composite Sample

Discrete Sediment Samples: A series of discrete sediment samples were collected at locations between transects to characterize individual reaches of the river and obtain good geographic coverage of the study area. Between three and 20 reach samples were taken between each transect. Reach sample locations were identified in the field and were preferentially located in depositional zones. Reach samples consisted of the upper one to two inches of sediment collected with a petite Ponar dredge. Stones larger than $\frac{1}{8}$ inch and leaves and twigs were removed from the sample before compositing in a stainless steel bowl. Reach sample locations are identified in Figure 3. Because of resource limitations and holding time limitations, reach samples were analyzed for dioxins, furans and physical parameters only.

Adjacent individual reach samples were carefully split and mixed in the lab to form a composite sample prior to analysis. A total of 19 reach *composite* samples were analyzed for dioxins and furans. The individual reach samples and the reach composites are shown on Figure 3 and listed in Table 3.

In addition to the composites, individual analyses were completed on each of the reach samples that make up Composites #4, #11, #15, and #17. These composites were selected for additional analysis based on the results of the H4IIE-luc bioassay as described in further detail below. An individual analysis was also performed on reach sample #23 which was distinguished from the other samples by a much higher TOC content.

Floodplain Soil Samples: Ten floodplain soil samples were also collected as part of the study. Low lying areas immediately adjacent to the river were selected, based on field judgment, as likely sites of significant fine particle deposition. The locations of the floodplain soil samples are identified in Figure 3. Latitudes and longitudes of floodplain soil sample locations are included in Table 2. Floodplain soil samples were collected from a one square foot location. The top one to two inches of soil was collected using a stainless steel scoop and thoroughly mixed in a stainless steel bowl. Large organic matter such as leaves and twigs were removed. Aliquots were taken for analysis of each of the study parameters listed in Table 1. A typical floodplain soil sampling location is shown in Figure 6.



Figure 6. Typical Floodplain Soil Sampling Location

Field Quality Control: As a quality control measure, duplicate samples were taken at river sediment location TF-C and floodplain soil location SS-7. These duplicate samples were analyzed for all study parameters.

Reference Locations: As the study area was the Tittabawassee River between the confluence of the Pine and Chippewa Rivers in the northwest to the Shiawassee River in the southeast, a transect core, composite, and

several discrete (reach) sediment and floodplain soil samples were taken above the confluence of the Tittabawassee and the Pine and the Tittabawassee and the Chippewa to serve as upstream controls. These upstream reference locations are listed in Table 4.

Additional Samples

Due to a broken chain-of-custody, dioxin and furan samples had to be recollected at transect sampling locations TC-C and TC-UC, and discrete reach sampling locations 1, 2, 3, and 11. Additional aliquots for the other study parameters were taken at the transect locations and serve as quality control replicates. The second set of samples collected from these locations is designated by an asterisk (1*, 2*, 3*, 11*, TC-C* and TC-UC*).

Analytical Methods

Analyses of sediments and flood plain soils at the selected locations were conducted using generally accepted sediment sampling and analytical techniques as identified below. Table 1 summarizes the analyses conducted on each sample and lists individual analytical parameters.

Volatile Organics: River sediment and floodplain soil samples for analysis of volatile organic compounds (VOCs) were methanol preserved in the field. Extraction was according to SW-846 Method 5035. Extracts were analyzed according to SW-846 Method 8260B. The list of analytes and detection limits is provided with the results in Table 5. Due to the lack of detections, no samples for VOCs were collected upstream of Transect F.

Semivolatile Organics: Semivolatile Organics (base neutral and acid extractable compounds and polynuclear aromatic hydrocarbons) were extracted using SW-846 Method 3510 and analyzed according to SW-846 Method 8270. The list of analytes and detection limits is provided with the results in Table 6.

Pesticides and PCBs: Pesticides and PCBs were extracted using SW-846 Method 3510 and analyzed according to SW-846 Method 8070. The list of analytes and detection limits is provided with the results in Table 7.

Metals: Samples were analyzed for metals using SW-846 6000 and 7000 series methods. The list of analytes and detection limits is provided with the results in Table 8.

Dioxins and Furans Chemical Analysis: Concentrations of seventeen 2,3,7,8-substituted polychlorinated dibenzo-p-dioxins (PCDDs) and

dibenzofurans (PCDFs) were analyzed by modifications of previously described methods (Yamashita et al., 2000; Kannan et al., 2001; Im et al., 2002). A copy of MSU-ATL's "Standard Operating Procedure for the Extraction and Analysis of 2,3,7,8-substituted PCDDs and PCDFs in Sediments using High Resolution Gas Chromatography & High Resolution Mass Spectrometry" is attached as Appendix 4 of this Report.

In Vitro Bioassay Analysis: H4IIE-luc bioassay was used to determine total dioxin-like activity (TCDD-EQs) in sediments. The bioassay procedures are described in detail in Appendix 5 of this Report, and in Villeneuve et al., 2000 and Hilscherova et al. 2001. Samples were tested as raw extracts and as acid-treated extracts using the *in vitro* H4IIE-luc recombinant cells for dioxin-like activity. A mass balance analysis of dioxin-like activity derived from instrumental and bioassay analyses was used to test for the presence of other dioxin-like compounds that can bind to the aromatic hydrocarbon receptor (AhR). Mass balance analysis (or potency balance analysis) was also used to examine whether or not the known composition of a sample (identified by instrumental analysis) can account for the magnitude or potency of biological response observed.

Total Organic Matter: Sediment and floodplain soil samples were analyzed for Total Organic Matter by the Michigan State University Soil Analysis Laboratory using their "Loss by Ignition" protocol.

Total Organic Carbon: Sediment and floodplain soil samples were analyzed for TOC by the Michigan State University Soil Analysis Laboratory using their "Dry Combustion using Microcarbon Analyzer" protocol.

Results

Results for VOCs, semivolatile organic compounds, pesticides, PCBs, and metals are presented in Tables 5, 6, 7 and 8, respectively. Geographic distribution of semivolatiles, PCBs, and pesticides are presented in Figure 7.

Dioxin and furan results are presented in Table 9. A full copy of the dioxin and furan analytical and bioassay results and quality assurance data is being retained by the MDEQ Waste Management Division. Table 9 also includes the results of the TOC and total organic matter analyses of these samples. The geographic distribution of dioxins and furans is presented in Figure 8.

Discussion

Volatiles:

Only one VOC was detected during this study. Tetrachloroethene was detected at 95 ppb at sample location TC-UC. Due to the flow regime of the Tittabawassee River, the lack of detectable volatile organic compounds is not surprising. The Tittabawassee is a shallow, fast moving river with considerable turbulence and scour during storm events. Volatile compounds, should they be released to the river, are not likely to persist in sediments that are frequently agitated. Given the lack of detects, the decision was made to cease sampling for VOCs above Transect F. None of the floodplain soil samples showed detectable levels of VOCs.

Semivolatiles:

Semivolatile organic compounds, when found, were typically present at very low concentrations. Semivolatile organic compounds, including phenanthrene, fluoranthene, pyrene, benzo(a)anthracene, and chrysene, were detected at sample locations TC-C, TF-CR, TE-C, TF-C, TF-UC, and TG-UC at levels below 400 ug/kg. Sample location TF-CR, in addition to having phenanthrene, fluoranthene, pyrene, and chrysene at very low levels, displayed hexachlorobenzene at 1300 ug/kg, a level that might indicate potential adverse impact to benthic organisms. Floodplain soil results were similar in displaying low levels of the same six semivolatile compounds at low levels (all below 510 ug/kg).

Pesticides and PCBs:

No PCBs were detected in sediment or floodplain soil samples. 4,4-DDT at 1,100 ug/kg and 4,4 DDD at 83 ug/kg in sediment sample TI-UC were the only target pesticides detected in sediment samples. Low levels of DDT and its breakdown products were detected at floodplain soil sampling locations SS-4 (Pine River), SS-5, and SS-7. There is a historical source of DDT on the Pine River that may have contributed to these detections. Hexabromobenzene was also detected at low levels at floodplain soil sampling locations SS-2, SS-4, and SS-7R. Detected levels of pesticides are not expected to have any significant impact on human health or the environment.

Metals:

Levels of metals in sediments are generally consistent with background levels. Of the floodplain soil samples, only SS-1 and SS-2 had concentrations that indicated the potential for minor aquatic life impacts (arsenic, chromium, copper, lead, mercury, nickel, and zinc). It is notable that SS-2 is immediately adjacent to and SS-1 is downstream of a former plate glass manufacturing facility which is known to have released metals to the river.

Dioxins and Furans:

Dioxins and furans were detected at each of the transect, reach, and individual sediment sampling locations analyzed during this study. Dioxins and furans were also present in each of the flood plain soil samples analyzed during this study. The concentrations of each of the 17 2,3,7,8-substituted PCDD and PCDF congeners, the calculated total toxic equivalent concentration (TEQ) relative to 2,3,7,8-TCDD, and the concentrations of the total tetra, penta, hexa, and hepta isomer groups are presented in Table 9 for each dioxin and furan sampling location.

The geographic distribution of the TEQ results is presented on Figure 8.

In general, the geographic distribution of dioxins and furans in the study area points to a source in the Midland area. The concentrations of dioxin in river sediments were less than 5 parts per trillion (ppt) TEQ in the seven reference samples that were taken above the junction of the Chippewa and Tittabawassee Rivers. Below the junction of the Chippewa and Tittabawassee Rivers, TEQ concentrations ranged from 5 to 2,000 ppt.

The concentrations of dioxins and furans were not significantly correlated with the TOC content of the samples. Correlation with the grain size of the samples with dioxin concentration was not completed in this study because the high levels of dioxins and furans in the samples resulted in an unanticipated health and safety concern during grain size analysis which involves drying and shaking of the samples. It is recommended that this evaluation be completed under the appropriate laboratory conditions.

Transect Results:

These samples are identified on Figure 8 by green boxes. The core composite samples are identified with a 'C' suffix (e.g. TA-C). The upper composite (dredge composite) samples are identified with a 'UC' suffix (e.g., TA-UC). The core sample results represent a mix of the surficial sediments and deeper sediments. The upper composite results represent only the top several inches of sediment.

In general, the concentrations of dioxins and furans were much higher in transect samples collected below the junction of the Tittabawassee and Chippewa Rivers than in the upstream reference samples. The transect reference samples were each less than 5 ppt total TEQ. Samples collected below the junction of the Tittabawassee River ranged in TEQ concentration from 5.1 ppt to 2000 ppt. It should be noted that the 5.1 ppt TEQ transect sample was collected at location TG-UC. No core sample could be collected from this location because of the coarse grained nature of the sediments. It is possible that the coarse grained nature of the sediments resulted in a relatively low concentration at this location.

The TEQ of the upper composite samples was similar to the core concentrations in most of the samples, with the exception of transects TC and TD where the concentration of dioxin and furans in the deeper cores was significantly higher than the upper composite samples. This suggests that in some parts of the study area, a reservoir of dioxin and furans may be present in deeper sediments. Further detailed coring work should be completed to determine if higher levels of dioxins and furans are present at lower depths in the river sediments.

Composite Results:

These samples show the distribution of dioxin and furans in the upper several inches of sediment in sediment accumulation areas in the study area. Due to resource limitations, adjacent sediment samples were carefully split and mixed in the lab to form a composite sample prior to analysis. The results of these analyses are shown on the maps as †Composite #1 through Composite #19.†

These results show that the composite concentrations are significantly elevated and quite variable below the junction of the Tittabawassee and Chippewa Rivers. This variability is probably reflective of the flow and depositional characteristics of the different segments of the river system.

In order to evaluate the variability within the composite locations, individual reach samples were analyzed at four locations in addition to the composite analyses. This analysis revealed that there can be considerable variability in the TEQ concentrations of the individual reach samples that were used to form a composite sample. In the case of Composite #15, the concentrations varied from 22 to 2000 ppt TEQ with a composite average analysis of 960 ppt. Composite #11 varied from 130 to 1000 ppt TEQ with a composite average analysis of 480 ppt TEQ. Based on the recognized variability, any future studies that are conducted to more fully understand of the distribution of dioxin contamination should avoid compositing samples.

Flood Plain Soil Samples:

Ten flood plain soil samples were taken as a part of this study. These samples are identified on Figure 8 by red hexagons. These samples were collected by scraping the upper one inch of soil from a one square foot area in selected low lying flood plain areas adjacent to the Tittabawassee River. Sample locations were also based on property access agreements.

Dioxin results in floodplain soils ranged from 2 to 11 ppt TEQ in control samples above the junction of the Chippewa and Tittabawassee Rivers. Below the junction, TEQ concentrations ranged from 300 to 1500 ppt. With the exception of SS#6, all of the samples were taken in the flood plain

within 100 feet of the edge of the river. SS#6 was taken from a recently tilled farm field several hundred yards away from the edge of the river.

The upstream reference sample results are consistent with Michigan's soil background dioxin levels. The soil concentrations below the confluence of the Tittabawassee and Chippewa are significantly elevated over Michigan's background soil concentration and exceed Michigan's Part 201, Environmental Remediation, of the Natural Resources and Environmental Protection Act, 1994 PA 451, as amended, generic residential soil protection criterion of 90 ppt TEQ.

The downstream soil concentrations also exceed the Agency for Toxic Substances and Disease Registry (ATSDR) interim policy guidelines for PCDDs/PCDFs in residential soils near or on hazardous waste sites (De Rosa et al., 1997). When concentrations of TEQs exceed 50 pg /g TEQ dry weight, the ATSDR recommends evaluation of site-specific factors such as pathway analysis and soil cover. When soil concentrations exceed 1000 pg/g TEQ dry weight, the ATSDR recommends health surveillance and exposure investigations.

Based on the results of this investigation and follow up work conducted by the MDEQ in 2002, additional work is needed to determine if the level of exposure in the flood plain is resulting in health and/or ecological impacts.

Bioassay and Mass Balance Results:

H4IIE-luc bioassay was used to determine total dioxin-like activity (TCDD-EQs) in sediments and to direct the instrumental analyses. A mass balance analysis of dioxin-like activity derived from instrumental and bioassay analyses was used to test for the presence of other dioxin-like compounds that can bind to the aromatic hydrocarbon receptor (AhR). Earlier studies have demonstrated that the H4IIE-luc bioassay coupled with instrumental analysis is useful in the integrated assessment of dioxin-like activity in sediment (Khim et al., 1999; Hilscherova et al., 2000; Khim et al., 2001).

The instrumental and bioanalytical approaches provide different and complementary information. While instrumental analysis is a useful tool to identify the compounds of interest and to evaluate the concentrations of environmental contaminants, it provides little information regarding the integrated biological relevance of a complex mixture of compounds associated with environmental samples such as sediment. Where appropriate, bioassay-directed fractionation and mass balance analysis is a powerful tool to characterize the causative agents responsible for bioassay responses observed. Recent studies have indicated that organic extracts of sediments elicit both dioxin-like responses significantly *in vitro*, although the chemical concentrations often did not explain the bioassay

activities observed (Khim et al., 1999; Kannan et al., 2000; Hilscherova et al., 2001). Empirical bioassay results and mass balance analyses can suggest the magnitude of contribution of target organic compounds to total dioxin-like activity of sediment extracts. Thus, the use of bioassay-based toxicity identification and evaluation (TIE) and mass balance analysis are important approach to assess sediment contamination since the sediment extracts may contain many potentially AhR-active compounds, which were not analyzed by instrumental methods.

The results of this study showed that the results of the bioassay analyses matched very well with the results of the instrumental analysis. Therefore, the bioassay technique proved valuable in determining which samples showed significant dioxin-like activity. The bioassay technique could be used to direct further investigation in the watershed by screening out low activity samples and identifying areas of high dioxin like activity that would warrant follow up chemical confirmation.

Appendix 5 contains the results of the bioassay analyses and the mass balance calculations. The results suggest that PCDDs/PCDFs, are the major sources of dioxin-like activity in sediments/soils from the Tittabawassee River and that little, if any, activity was due to other compounds, such as PCBs, that exhibit dioxin-like activity. This was further supported by the lack of detectable concentrations of PCBs in soils and sediments.

Sources of Dioxins and Furans in the Tittabawassee River Watershed:

Figures 9, 10, 11, and 12 are congener profiles of the dioxin and furan samples collected during this study. The TEQ concentration of each sample has been normalized to 100 percent (%) and the normalized toxic equivalent concentration of each congener is plotted on the resulting bar chart. Congeners that were not detected at a specific sampling location were assigned a value of zero. The resulting bar chart is a †fingerprint† of the 17 dioxins and furans that exhibit dioxin-like toxicity. The upstream reference samples are plotted at the top of each of the graphs and are marked with an asterisk.

As can be seen by review of the graphs, all of the samples taken from below the confluence of the Chippewa and Tittabawassee Rivers are fairly similar to each other and markedly different from the upstream reference samples. The chemical fingerprint of the sediment samples is similar to the chemical fingerprint of the floodplain soil samples, indicating that the dioxins and furans in these media are likely from the same source(s).

The bulk of the dioxin-like toxicity in the downstream samples is contributed by furan congeners. 2,3,7,8-TCDF and 2,3,4,7,8-PeCDF

contribute the bulk of the dioxin-like toxicity in the study area downstream of the confluence of the Tittabawassee and Chippewa Rivers. 1,2,3,7,8- \textgreekpi PeCDF and 1,2,3,4,7,8-HxCDF also make significant contributions to the dioxin-like toxicity of downstream sediment and soil samples. 2,3,7,8-TCDD, the most potent dioxin-like compound, typically contributes less than five percent of the TEQ in the downstream samples.

Different sources of PCDDs/PCDFs are characterized by different congener and homologue patterns (Kannan et al., 1998). Furthermore, differences in the physicochemical (mobility, solubility, etc.) and biological (biodegradation, bioaccumulation, etc.) properties may alter the congener profiles.

The fingerprints of PCDD and PCDF congeners in sediments collected from the Tittabawassee River downstream of Midland are all similar, suggesting the presence of a single major source. As noted above, the pattern of relative concentrations of PCDD/PCDF congeners was also different in soils and sediment collected downstream of Midland than in those collected upstream of the reference locations. A large proportion of OCDD and HpCDD has been suggested to be due to the sources originating from chlorophenol-related sources (Masunaga et al., 2001). Greater proportions of TCDFs suggest sources originating from PCB mixtures, chlorobenzenes, chlor-alkali processes, and incineration of PCBs and polyvinyl chloride (Wakimoto et al., 1988; Masunaga et al., 2001; Swami et al., 1992; Kannan et al., 1998). Total concentrations of PCBs in sediments from the Tittabawassee River were less than 150 nanograms/gram. This suggests that PCBs are not the source of the PCDD/PCDFs, but rather other sources such as chlorophenol and chlorobenzene production, incineration, or chlor-alkali processes are the sources of the PCDFs found in Tittabawassee River sediments collected below Midland.

The Dow Chemical Company (Dow) has a long history as a major manufacturer of chlorobenzenes, chlorophenols, and chlor-alkali products in Midland, Michigan. Additionally, Dow has conducted chemical waste incineration for many years. The geographic distribution of the contaminants combined with the dioxin and furan congener profile information strongly suggests that Dow's Midland facility is the most likely source of the elevated levels of dioxins and furans in the Tittabawassee River.

Conclusions and Recommendations

Summary: The data reviewed indicates that the potential for impacts to aquatic life to occur from the sediment chemicals analyzed, other than dioxins and furans, is minimal.

With regard to dioxins and furans, the following conclusions can be drawn:

1. The concentrations of dioxins and furans in sediments and soils represent a potential environmental and human health issue in the Tittabawassee River watershed that requires further study.
2. In the study area, the bulk of the dioxin-like toxicity in sediments and soils is contributed by the polychlorinated dibenzo-p-dioxins and furans. In particular, 2,3,7,8-TCDF and 2,3,4,7,8-PeCDF contribute the bulk of the dioxin-like toxicity to the samples collected from below the confluence of the Tittabawassee and Chippewa Rivers. Little dioxin-like toxicity appears to be contributed by other compounds that exhibit dioxin-like activity, such as PCBs.
3. The downstream extent of dioxin and furan contamination in sediments and floodplain soils has not been completely defined.
4. Deeper sediment cores are needed to determine the vertical extent of sediment contamination in the Tittabawassee and downstream in the Saginaw River.
5. The distribution of dioxins and furans in the study area sediments appears heterogeneous. This is most likely related to the flow regime of the Tittabawassee River. Any follow up sampling that is conducted should not involve compositing sediment samples. The relationship of grain size to dioxin and furan concentrations in the study area needs to be investigated.
6. The H4IIE-luc bioassay technique can be successfully used as a tool to direct further investigation of dioxin and furan contamination in the study area.
7. The most probable historic source of dioxins and furans in the Tittabawassee River watershed, based on the geographic distribution of the contaminants and the chemical profiles of the dioxin and furan congeners, is located in Midland, Michigan.

Recommendations:

Based on these conclusions, additional sampling is recommended to more completely define the horizontal and vertical distribution of dioxins and furans in the Tittabawassee River and Saginaw River watersheds. Periodic resampling should be conducted to determine if concentrations

are changing over time. The existing information should be used along with other available information to conduct an assessment of the risk to human health and the environment from the elevated concentrations of dioxins and furans present in the study area.

TABLES

TABLE 9

Part 1										Part 2										Part 3														
Analyte	TB-UC					TC-C					TC-UC																							
	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero														
2378-TCDD	1	1.20	1.2000	1.2000	ND	0.92	0.9200	0.9200	ND	0.50	0.2500	0.1250	0.0000	0.0000	ND	0.54	0.270	0.270	0.2700															
12378-PeCDD	0.5	ND	0.50	0.2500	0.1250	0.0000	ND	0.50	0.2500	0.1250	0.0000	0.0000	0.0000	0.0000	ND	10.54	1.054	1.054	1.0540															
123478-HxCDD	0.1	ND	10.41	1.0410	1.0410	0.0410	13.32	1.3320	1.3320	1.3320	1.3320	1.3320	1.3320	1.3320	ND	0.50	0.050	0.050	0.0000															
123678-HxCDD	0.1	ND	3.44	0.3440	0.3440	0.0340	1.43	0.1430	0.1430	0.1430	0.1430	0.1430	0.1430	0.1430	ND	0.50	0.050	0.050	0.0000															
123789-HxCDD	0.1	ND	0.50	0.0500	0.0250	0.0000	ND	0.50	0.0500	0.0250	0.0000	0.0000	0.0000	0.0000	ND	0.50	0.050	0.050	0.0000															
1234678-HpCDD	0.01	53.56	0.5356	0.5356	0.5356	0.5356	31.74	0.3174	0.3174	0.3174	0.3174	0.3174	0.3174	0.3174	ND	23.64	0.2336	0.2336	0.23364															
12346789-OCDD	0.001	483.51	0.4835	0.4835	0.4835	0.4835	208.19	0.2082	0.2082	0.2082	0.2082	0.2082	0.2082	0.2082	ND	186.10	0.1886	0.1886	0.1861															
23781CDF	0.1	547.68	54.7680	54.7680	54.7680	54.7680	33.80	3.3800	3.3800	3.3800	3.3800	3.3800	3.3800	3.3800	ND	162.20	16.220	16.220	16.2200															
12378-PeCDF	0.05	198.27	9.9135	9.9135	9.9135	9.9135	14.27	0.7135	0.7135	0.7135	0.7135	0.7135	0.7135	0.7135	ND	96.44	4.822	4.822	4.8220															
23478-PeCDF	0.5	130.15	65.0750	65.0750	65.0750	65.0750	8.26	4.1300	4.1300	4.1300	4.1300	4.1300	4.1300	4.1300	ND	52.75	26.375	26.375	26.3750															
123478-HxCDF	0.1	90.04	9.0040	9.0040	9.0040	9.0040	10.01	1.0010	1.0010	1.0010	1.0010	1.0010	1.0010	1.0010	ND	54.51	5.451	5.451	5.4510															
123678-HxCDF	0.1	23.34	2.3340	2.3340	2.3340	2.3340	1.54	0.1540	0.1540	0.1540	0.1540	0.1540	0.1540	0.1540	ND	12.66	1.266	1.266	1.2660															
234678-HxCDF	0.1	10.21	1.0210	1.0210	1.0210	1.0210	ND	0.50	0.0500	0.0250	0.0250	0.0250	0.0250	0.0250	ND	4.85	0.485	0.485	0.4850															
123789-HxCDF	0.1	0.87	0.0870	0.0870	0.0870	0.0870	ND	0.50	0.0500	0.0250	0.0250	0.0250	0.0250	0.0250	ND	0.50	0.050	0.050	0.0000															
1234678-HpCDF	0.01	91.15	0.9115	0.9115	0.9115	0.9115	43.12	0.4312	0.4312	0.4312	0.4312	0.4312	0.4312	0.4312	ND	60.70	0.607	0.607	0.6070															
1234789-HpCDF	0.01	8.37	0.0837	0.0837	0.0837	0.0837	4.05	0.0405	0.0405	0.0405	0.0405	0.0405	0.0405	0.0405	ND	69.57	0.6996	0.6996	0.6957															
12346789-OCDF	0.001	88.81	0.0888	0.0888	0.0888	0.0888	55.54	0.0555	0.0555	0.0555	0.0555	0.0555	0.0555	0.0555	ND	39.12	0.039	0.039	0.0391															
nondetect = detection limit					TEQ= 147.1906					TEQ= 13.2263					TEQ= 59.4573																			
nondetect = 1/2 d.l.					teq= 147.0406					teq= 13.0263					teq= 59.3823																			
nondetect = zero					teq=					teq=					teq=																			
Moisture content (%)					18.59					18.64					20.77																			
Organic carbon (%)					0.26					0.18					0.33																			
Total organic matter (%)					0.45					0.31					0.57																			
Sample ID.					TB-UC					TC-C					TC-UC																			
tetra-PCDF (TCDF)					548.0					33.8					162.0																			
penta-PCDF (PeCDF)					328.0					22.5					149.0																			
hexa-PCDF (HxCDF)					124.0					11.6					72.0																			
hepta-PCDF (HpCDF)					99.5					47.2					130.0																			
octa-PCDF (OCDF)					88.8					55.5					39.1																			
tetra-PCDD (TCDD)					1.2					0.9					1.6																			
penta-PCDD (PeCDD)					ND					ND					0.5																			
hexa-PCDD (HxCDD)					13.9					14.8					10.5																			
hepta-PCDD (HpCDD)					53.6					31.7					23.6																			
octa-PCDD (OCDD)					484.0					208.0					186.0																			
Summary concentrations of the homologs																																		
Concentrations of all compound (except of Recalculations for internal standards account)																																		
* OCDF concentrations were calculated based if required. OCDF concentrations can be non-detect, the detection limits will																																		

TABLE 9

Part 1		Part 1												Part 1																																			
		TD-C				TD-JUC				TE-C																																							
Analyte	TEF	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero																								
2378-TCDD	1	1.56	1.560	1.560		0.97	0.970	0.970	0.9700					1.95	1.950																																		
12378-PeCDD	0.5	1.03	0.515	0.515	0.515	0.60	0.300	0.300	0.3000					0.74	0.370	0.370	0.3700																																
123478-HxCDD	0.1	32.32	3.232	3.232	3.232	1.44	0.144	0.144	0.1440					63.47	6.347	6.347	6.3470																																
123678-HxCDD	0.1	2.06	0.206	0.206	0.206	ND	1.20	0.120	0.060	0.0000				14.68	1.468	1.468	1.4680																																
123789-HxCDD	0.1	2.11	0.211	0.211	0.211	ND	1.20	0.120	0.060	0.0000				3.34	0.334	0.334	0.3340																																
1234678-HpCDD	0.01	131.46	1.315	1.315	1.315		25.25	0.253	0.253	0.2525				207.34	2.073	2.073	2.0734																																
12346789-OCDD	0.001	911.49	0.911	0.911	0.911		287.56	0.288	0.288	0.2876				2139.70	2.140	2.140	2.1397																																
23781CDF	0.1	308.98	308.498	308.498	308.498	267.47	26.747	26.747	26.7470					257.795	257.795	257.795	257.7950																																
12378-PeCDF	0.05	4891.86	244.593	244.593	244.593	139.34	6.967	6.967	6.9670					1496.96	74.848	74.848	74.8480																																
23478-PeCDF	0.5	1660.30	830.150	830.150	830.150	91.78	45.890	45.890	45.8900					980.42	490.210	490.210	490.2100																																
123478-HxCDF	0.1	5020.21	502.021	502.021	502.021	49.60	4.960	4.960	4.9600					663.00	66.300	66.300	66.3000																																
123678-HxCDF	0.1	1014.95	101.495	101.495	101.495	10.06	1.006	1.006	1.0060					177.19	17.719	17.719	17.7190																																
234678-HxCDF	0.1	219.77	21.977	21.977	21.977	4.91	0.491	0.491	0.4910					80.29	8.029	8.029	8.0290																																
123789-HxCDF	0.1	63.91	6.391	6.391	6.391	1.10	0.110	0.110	0.1100					5.22	0.522	0.522	0.5220																																
1234678-HpCDF	0.01	844.04	8.440	8.440	8.440	53.23	0.532	0.532	0.5323					436.54	4.365	4.365	4.3654																																
1234789-HpCDF	0.01	346.97	3.470	3.470	3.470	7.37	0.074	0.074	0.0737					52.29	0.523	0.523	0.5229																																
12346789-OCDF	0.001	594.49	0.594	0.594	0.594	38.01	0.038	0.038	0.0380					485.02	0.485	0.485	0.4850																																
nondetect = detection limit		TEQ= 2035.5797				TEQ= 89.0091				TEQ= 935.4784																																							
nondetect = 1/2 d.l.		teq= 2035.5797				teq= 88.8891				teq= 935.4784																																							
nondetect = zero		teq= teq= teq= teq=				teq= teq= teq= teq=				teq= teq= teq= teq=																																							
Moisture content (%)		17.82				24.44				33.27																																							
Organic carbon (%)		0.12				0.33				1.27																																							
Total organic matter (%)		0.21				0.57				2.19																																							
Sample ID.		TD-C				TD-JUC				TE-C																																							
tetra-PCDF (TCDF)		3085.0				267.0				2578.0																																							
penta-PCDF (PeCDF)		6552.0				231.0				2477.0																																							
hexa-PCDF (HxCDF)		6319.0				65.7				926.0																																							
hepta-PCDF (HpCDF)		1191.0				60.6				489.0																																							
octa-PCDF (OCDF)		594.0				38.0				485.0																																							
tetra-PCDD (TCDD)		1.6				1.0				1.9																																							
penta-PCDD (PeCDD)		1.0				0.6				0.7																																							
hexa-PCDD (HxCDD)		36.5				1.4				81.5																																							
hepta-PCDD (HpCDD)		131.0				25.2				207.0																																							
octa-PCDD (OCDD)		911.0				288.0				2140.0																																							
Summary concentrations of the homologs																																																	
Concentrations of all compound (except of Recalculations for internal standards account)																																																	
* OCDF concentrations were calculated based if required. OCDF concentrations can be non-detect, the detection limits will																																																	

TABLE 9
Part I

TABLE 9

Part 1										Part 2										Part 3									
Analyte	TEF			TF-UC			TG-UC			TH-C			TEF			TF-UC			TG-UC			TH-C							
	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	sampled pg/g	toxic eq. pg/g	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	sampled pg/g	toxic eq. pg/g	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.								
2378-TCDD	1	2.56	2.560	2.5600	ND	0.50	0.500	0.2500	0.0000	ND	0.500	0.500	0.2500	0.0000	0.0000	ND	0.500	0.500	0.2500	0.0000	0.0000								
12378-PeCDD	0.5	ND	0.50	0.250	0.1250	0.0000	ND	0.50	0.250	0.1250	0.0000	ND	0.500	0.500	0.2500	0.1250	0.1250	0.0000	0.0000	0.0000	0.0000								
123478-HxCDD	0.1	24.71	2.471	2.4710	2.4710	ND	0.92	0.092	0.0920	0.0920	ND	0.920	0.0920	0.0920	0.0920	ND	0.500	0.050	0.0250	0.0250	0.0250								
123678-HxCDD	0.1	9.13	0.913	0.9130	0.9130	ND	5.85	0.585	0.5850	0.5850	ND	5.850	0.5850	0.5850	0.5850	ND	0.500	0.050	0.0250	0.0250	0.0250								
123789-HxCDD	0.1	4.61	0.461	0.4610	0.4610	ND	0.50	0.050	0.0250	0.0250	ND	0.500	0.050	0.0250	0.0250	ND	0.500	0.050	0.0250	0.0250	0.0250								
1234678-HpCDD	0.01	121.15	1.212	1.2115	1.2115	ND	31.80	0.318	0.3180	0.3180	ND	31.80	0.318	0.3180	0.3180	ND	6.382	0.064	0.0638	0.0638	0.0638								
12346789-OCDD	0.001	1099.11	1.099	1.0991	1.0991	ND	224.25	0.2243	0.2243	0.2243	ND	224.25	0.2243	0.2243	0.2243	ND	52.715	0.053	0.0527	0.0527	0.0527								
23781CDF	0.1	1718.24	171.824	171.8240	171.8240	ND	8.96	0.896	0.8960	0.8960	ND	8.960	0.8960	0.8960	0.8960	ND	2.790	0.279	0.2790	0.2790	0.2790								
12378-PeCDF	0.05	959.43	47.972	47.9715	47.9715	ND	5.28	0.264	0.2640	0.2640	ND	5.280	0.264	0.2640	0.2640	ND	1.346	0.067	0.0673	0.0673	0.0673								
23478-PeCDF	0.5	507.40	253.700	253.7000	253.7000	ND	2.52	1.260	1.2600	1.2600	ND	2.520	1.260	1.2600	1.2600	ND	0.500	0.250	0.1250	0.1250	0.1250								
123478-HxCDF	0.1	428.50	42.850	42.8500	42.8500	ND	6.20	0.620	0.6200	0.6200	ND	6.200	0.620	0.6200	0.6200	ND	2.430	0.243	0.2430	0.2430	0.2430								
123678-HxCDF	0.1	90.64	9.0640	9.0640	9.0640	ND	0.78	0.078	0.0780	0.0780	ND	0.780	0.078	0.0780	0.0780	ND	0.500	0.050	0.0250	0.0250	0.0250								
234678-HxCDF	0.1	33.59	3.359	3.3590	3.3590	ND	0.50	0.050	0.0250	0.0250	ND	0.500	0.050	0.0250	0.0250	ND	0.500	0.050	0.0250	0.0250	0.0250								
123789-HxCDF	0.1	3.59	0.359	0.3590	0.3590	ND	0.50	0.050	0.0250	0.0250	ND	0.500	0.050	0.0250	0.0250	ND	0.500	0.050	0.0250	0.0250	0.0250								
1234678-HpCDF	0.01	234.05	2.341	2.3405	2.3405	ND	30.42	0.304	0.3042	0.3042	ND	30.420	0.304	0.3042	0.3042	ND	3.730	0.037	0.0373	0.0373	0.0373								
1234789-HpCDF	0.01	27.71	0.277	0.2771	0.2771	ND	2.59	0.026	0.0259	0.0259	ND	2.590	0.026	0.0259	0.0259	ND	0.640	0.006	0.0064	0.0064	0.0064								
12346789-OCDF	0.001	356.53	0.357	0.3565	0.3565	ND	31.31	0.031	0.0313	0.0313	ND	31.310	0.031	0.0313	0.0313	ND	2.953	0.003	0.0030	0.0030	0.0030								
nondetect = detection limit										TEQ = 5.5987										TEQ = 2.0525									
nondetect = 1/2 d.l.										teq = 5.1487										teq = 1.4025									
nondetect = zero										teq = 4.6987										teq = 0.7525									
Moisture content (%)										23.9										25.18									
Organic carbon (%)										0.33										0.26									
Total organic matter (%)										0.57										0.45									
Sample ID.										TF-UC										TG-UC									
tetra-PCDF (TCDF)										1718.0										9.0									
penta-PCDF (PeCDF)										1467.0										7.8									
hexa-PCDF (HxCDF)										556.0										7.0									
hepta-PCDF (HpCDF)										2620										33.0									
octa-PCDF (OCDF)										357.0										31.3									
tetra-PCDD (TCDD)										2.6										ND									
penta-PCDD (PeCDD)										ND										0.5									
hexa-PCDD (HxCDD)										38.4										6.8									
hepta-PCDD (HpCDD)										121.0										31.8									
octa-PCDD (OCDD)										1099.0										224.0									
Summary concentrations of the homologs										Concentrations of all compound (except of internal standards account)										Recalculations for internal standards account									
* OCDF concentrations were calculated based on detection limits if required.										if required, OCDF concentrations can be non-detect, the detection limits will be zero.										ND = non detected, the detection limits will be zero.									

TABLE 9

Part 1										Part 2											
Analyte	TH-UC					Ti-C					Ti-UC										
	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	
2378-TCDD	1	ND	0.50	0.2500	0.0000	ND	1.23	1.230	1.2300	ND	0.50	0.500	0.2500	0.0000	ND	0.50	0.500	0.2500	0.0000		
12378-PeCDD	0.5	ND	0.50	0.250	0.1250	0.0000	ND	0.50	0.250	0.1250	0.0000	ND	0.50	0.250	0.1250	0.0000	ND	0.50	0.250	0.1250	
123478-HxCDD	0.1	ND	0.50	0.050	0.0250	0.0000	ND	0.50	0.050	0.0250	0.0000	ND	0.50	0.050	0.0250	0.0000	ND	0.50	0.050	0.0250	
123678-HxCDD	0.1	ND	0.50	0.050	0.0250	0.0000	ND	0.50	0.050	0.0250	0.0000	ND	0.50	0.050	0.0250	0.0000	ND	0.50	0.050	0.0250	
123789-HxCDD	0.1	ND	0.50	0.050	0.0250	0.0000	ND	0.50	0.050	0.0250	0.0000	ND	0.60	0.060	0.0300	0.0000	ND	0.60	0.060	0.0300	
1234678-HpCDD	0.01		3.25	0.033	0.0325	0.0325		20.56	0.206	0.2056	0.2056		11.17	0.112	0.1117	0.1117		11.17	0.112	0.1117	
12346789-OCDD	0.001		31.55	0.032	0.0316	0.0316		178.72	0.179	0.1787	0.1787		82.66	0.083	0.0827	0.0827		82.66	0.083	0.0827	
23781CDF	0.1		1.01	0.101	0.1010	0.1010		7.18	0.718	0.7180	0.7180		2.42	0.242	0.2420	0.2420		2.42	0.242	0.2420	
12378-PeCDF	0.05		0.73	0.037	0.0365	0.0365		2.22	0.111	0.1110	0.1110		1.99	0.100	0.0995	0.0995		1.99	0.100	0.0995	
23478-PeCDF	0.5		ND	0.50	0.250	0.1250		2.87	1.435	1.4350	1.4350		1.49	0.745	0.7450	0.7450		1.49	0.745	0.7450	
123478-HxCDF	0.1		1.42	0.142	0.1420	0.1420		3.16	0.316	0.3160	0.3160		1.16	0.116	0.1160	0.1160		1.16	0.116	0.1160	
123678-HxCDF	0.1		ND	0.50	0.050	0.0250		1.21	0.121	0.1210	0.1210		ND	0.50	0.050	0.0250		ND	0.50	0.050	0.0250
234678-HxCDF	0.1		ND	0.50	0.050	0.0250		ND	0.50	0.050	0.0250		ND	0.50	0.050	0.0250		ND	0.50	0.050	0.0250
123789-HxCDF	0.1		ND	0.50	0.050	0.0250		ND	0.50	0.050	0.0250		ND	0.50	0.050	0.0250		ND	0.50	0.050	0.0250
1234678-HpCDF	0.01		2.90	0.029	0.0290	0.0290		17.90	0.179	0.1790	0.1790		6.32	0.063	0.0632	0.0632		6.32	0.063	0.0632	
1234789-HpCDF	0.01		0.53	0.005	0.0053	0.0053		1.02	0.010	0.0102	0.0102		0.82	0.008	0.0082	0.0082		0.82	0.008	0.0082	
12346789-OCDF	0.001		2.61	0.003	0.0026	0.0026		11.28	0.011	0.0113	0.0113		4.18	0.004	0.0042	0.0042		4.18	0.004	0.0042	
nondetects = detection limit										TEQ = 5.0158											
nondetects = 1/2 d.l.										TEQ = 4.7908											
nondetects = zero										TEQ = 4.5658											
Moisture content (%)										36.61											
Organic carbon (%)										1.28											
Total organic matter (%)										2.21											
Sample ID.										Ti-C											
TH-UC										Ti-UC											
tetra-PCDF (TCDF)										7.2											
penta-PCDF (PeCDF)										5.1											
hexa-PCDF (HxCDF)										4.4											
hepta-PCDF (HpCDF)										18.9											
octa-PCDF (OCDF)										11.3											
tetra-PCDD (TCDD)										1.2											
penta-PCDD (PeCDD)										ND											
hexa-PCDD (HxCDD)										0.5											
hepta-PCDD (HpCDD)										20.6											
octa-PCDD (OCDD)										179.0											
Summary concentrations of the homologs																					
Concentrations of all compound (except of Recalculations for internal standards account)																					
* OCDF concentrations were calculated based if required. OCDF concentrations can be non-detect, the detection limits will																					
ND = non detected, the detection limits will																					

TABLE 9

Part 2

Part 2 (continued)

Analyte	TEF	Part 2												Part 2												
		C1				C2				C3				C1				C2				C3				
		sampled pg/g	toxic eq. pg/g	nondetect	nondetect zero		sampled pg/g	toxic eq. pg/g	nondetect	nondetect zero		sampled pg/g	toxic eq. pg/g	nondetect	nondetect zero		sampled pg/g	toxic eq. pg/g	nondetect	nondetect zero		sampled pg/g	toxic eq. pg/g	nondetect	nondetect zero	
2378-TcDD	1	1.52	1.520	1.5200	1.5200		2.27	2.270	2.2700	2.2700		3.13	3.130	3.1300	3.1300		3.13	3.130	3.1300	3.1300		3.13	3.130	3.1300	3.1300	
12378-PeCDD	0.5	ND	0.50	0.250	0.1250	0.0000	ND	0.50	0.250	0.1250	0.0000	0.98	0.490	0.4900	0.4900	0.4900	0.98	0.490	0.4900	0.4900	0.4900	0.98	0.490	0.4900	0.4900	
123478-HxCDD	0.1	45.03	4.503	4.5030	4.5030		17.28	1.728	1.7280	1.7280		63.20	6.320	6.3200	6.3200		13.87	1.387	1.3870	1.3870		13.87	1.387	1.3870	1.3870	
123678-HxCDD	0.1	4.86	0.486	0.4860	0.4860		6.07	0.607	0.6070	0.6070		0.6070	0.6070	0.6070	0.6070		0.1140	0.1140	0.1140	0.1140		0.344	0.344	0.3440	0.3440	
1234678-HxCDD	0.1	0.50	0.050	0.0250	0.0000		1.14	0.114	0.1140	0.1140		0.6090	0.6090	0.6090	0.6090		0.6090	0.6090	0.6090	0.6090		1.4134	1.4134	1.4134	1.4134	
12346789-OCDD	0.001	78.44	0.784	0.7844	0.7844		60.90	0.609	0.6090	0.6090		476.34	0.4763	0.4763	0.4763		1100.78	1.100	1.1008	1.1008		1100.78	1.100	1.1008	1.1008	
2378TCDF	0.1	659.09	0.659	0.6591	0.6591		3600.50	360.050	360.0500	360.0500		3600.50	360.050	360.0500	360.0500		747.63	74.763	74.7630	74.7630		747.63	74.763	74.7630	74.7630	
12378-PeCDF	0.05	625.06	62.506	62.5060	62.5060		1145.95	57.298	57.2975	57.2975		411.60	20.580	20.5800	20.5800		812.82	406.410	406.4100	406.4100		235.92	117.960	117.9600	117.9600	
23478-PeCDF	0.5	211.16	105.580	105.5800	105.5800		359.52	35.952	35.9520	35.9520		16.382	1.6.382	1.6.3820	1.6.3820		19.50	0.195	0.1950	0.1950		28.17	0.282	0.2817	0.2817	
123478-HxCDF	0.1	160.78	16.078	16.0780	16.0780		198.13	0.198	0.1981	0.1981		0.1981	0.1981	0.1981	0.1981		0.1981	0.1981	0.1981	0.1981		267.76	0.268	0.2678	0.2678	
123678-HxCDF	0.1	32.84	3.284	3.2840	3.2840		93.16	9.316	9.3160	9.3160		30.18	3.018	3.0180	3.0180		43.55	4.355	4.3550	4.3550		11.48	1.148	1.1480	1.1480	
1234678-HxCDF	0.1	11.68	1.168	1.1680	1.1680		8.64	0.864	0.8640	0.8640		3.01	0.301	0.3010	0.3010		2.544	0.254	0.2540	0.2540		147.40	1.4740	1.4740	1.4740	
123789-HxCDF	0.1	2.54	0.254	0.2540	0.2540		1.4740	1.4740	1.4740	1.4740		351.97	3.520	3.5197	3.5197		13.89	0.139	0.1389	0.1389		19.50	0.195	0.1950	0.1950	
12346789-HxCDF	0.001	155.26	0.155	0.1553	0.1553		198.13	0.198	0.1981	0.1981		21.74	2.227	2.2270	2.2270		0.37	0.36	0.36	0.36		27.11	0.94	0.94	0.94	
12346789-OCDF	0.001	TEQ=	217.65	TEQ=	217.50	TEQ=	882.17	TEQ=	882.04	TEQ=	881.92	TEQ=	881.92	TEQ=	881.92	TEQ=	881.92	TEQ=	881.92	TEQ=	881.92	TEQ=	881.92	TEQ=	881.92	
nondetects = detection limit																										
nondetects = 1/2 d.l.																										
nondetects = zero																										
Moisture content (%)																										
Organic carbon (%)																										
Total organic matter (%)																										
Sample ID			C-1									C-2									C-3					
tetra-PCDF (TCDF)			625																							
penta-PCDF (PeCDF)			565																							
hexa-PCDF (HxCDF)			208																							
hepta-PCDF (HpCDF)			268																							
octa-PCDF (OCDF)			155																							
tetra-PCDD (TCDD)			1.5																							
pentra-PCDD (PeCDD)			ND																							
hexa-PCDD (HxCDD)			49.9																							
hepta-PCDD (HpCDD)			78.4																							
octa-PCDD (OCDD)			659																							

Summary concentrations of the homolog groups [pg/g dry weight]

Concentrations of all compound (except of OCDF) are calculated based on internal standard calibration

Recalculation for internal standards accounts for the recoveries

* OCDF concentrations were calculated based on external standard and they were not recalculated for recoveries since no specific internal standard (no 13C-OCDF) was available

if required, OCDF concentrations can be normalized for 13C-OCDF recoveries

ND = non detected, the detection limits will be specified on individual sample bases

TABLE 9

Part 2

Part 2 (continued)

Analyte	TEF	C-4		C-5		C-6	
		sampled pg/g	toxic eq. pg/g	nondetect I/2 d.l.	sampled pg/g	toxic eq. pg/g	nondetect I/2 d.l.
2378-TCDD	1	1.54	1.540	1.5400	1.570	1.5700	1.74
12378-PeCDD	0.5	ND	0.50	0.1250	0.0000	4.45	2.2250
123478-HxCDD	0.1	5.325	5.3250	5.3250	23.59	2.3590	32.64
123678-HxCDF	0.1	14.47	1.447	1.4470	20.19	2.0190	2.0190
123789-HxCDD	0.1	3.29	0.329	0.3290	5.66	0.5660	2.09
1234678-HxCDD	0.01	149.46	1.495	1.4946	95.30	0.953	95.18
12346789-OCDDD	0.001	1281.52	1.282	1.2815	715.05	0.715	715.11
2378TCDF	0.1	5449.06	544.9060	544.9060	1025.88	102.5880	102.5880
12378-PeCDF	0.05	2261.76	113.088	113.0880	1064.92	53.2446	53.2446
23478-PeCDF	0.5	1617.75	808.875	808.8750	540.82	270.4100	270.4100
123478-HxCDF	0.1	895.98	89.598	89.5980	551.68	55.1680	55.1680
123678-HxCDF	0.1	160.95	16.095	16.0950	95.57	9.5570	9.5570
234678-HxCDF	0.1	97.50	9.750	9.7500	50.15	5.0150	5.0150
123789-HxCDF	0.1	5.74	0.574	0.5740	6.83	0.6830	0.6830
1234678-HxCDF	0.01	388.22	3.882	3.8822	255.26	2.5526	2.5526
1234789-HxCDF	0.01	45.92	0.459	0.4592	39.16	0.3932	0.3916
12346789-OCDF	0.001	450.79	0.451	0.4508	139.79	0.140	0.1398
nondetects = detection limit		TEQ= 1599.35		TEQ= 510.16		TEQ= 44.01	
nondetects = I/2 d.l.		teq= teq= 1599.22		teq= 510.16		teq= 4.4010	
nondetects = zero		teq= teq= 1599.10		teq= 510.16		teq= 4.4010	
Moisture content (%)							
Organic carbon (%)		22.1		24.59		24.59	
Total organic matter (%)		0.66		0.38		0.38	
		1.13		0.66		0.66	
Sample I.D.							
		C-4		C-5		C-6	
tetra-PCDF (TCDF)		5449		1026		1717	
penta-PCDF (PeCDF)		3880		1606		888	
hexa-PCDF (HxCDF)		1160		704		297	
hepta-PCDF (HpCDF)		434		294		245	
octa-PCDF (OCDF)		451		140		183	
tetra-PCDD (TCDD)		1.5		1.6		1.7	
pentra-PCDD (PeCDD)	ND	0.5		4.5		0.9	
hexa-PCDD (HxCDD)		71		49.4		43.4	
hepta-PCDD (HpCDD)		149		95.3		95.2	
octa-PCDD (OCDD)		1282		715		974	
Summary concentrations of the homologs							
Concentrations of all compound (except of internal standards account)							
Recalculation for internal standards account							
* OCDF concentrations were calculated based on required OCDF concentrations can be non detected, the detection limits will be							

TABLE 9

Part 2

Part 2 (continued)

Analyte	TEF	C-7		C-8		C-9												
		sampled pg/g	toxic eq. pg/g	nondetect I/2 d.l.	nondetect sampled pg/g	toxic eq. pg/g	nondetect I/2 d.l.	nondetect sampled pg/g	toxic eq. pg/g	nondetect I/2 d.l.								
2378-TcDD	1	4.67	4.6700	4.6700	ND	1.40	1.4000	0.7000	0.0000	2.9900								
12378-PeCDD	0.5	4.27	2.1350	2.1350	2.21	1.1050	1.1050	ND	0.75	0.3750								
12378-HxCDD	0.1	103.43	10.3430	10.3430	64.35	6.4350	6.4350	6.4350	30.09	19.33								
123678-HxCDD	0.1	26.09	2.6090	2.6090	3.25	0.3250	0.3250	0.3250	2.20	1.9330								
123789-HxCDD	0.1	7.11	0.7110	0.7110	2.29	0.2290	0.2290	0.2290	0.2200	0.2200								
1234678-HxCDD	0.01	271.62	2.7162	2.7162	234.30	2.3430	2.3430	2.3430	186.48	1.8648								
12346789-OCDD	0.001	2943.76	2.9438	2.9438	1645.38	1.6454	1.6454	1.6454	1540.70	1.5407								
2378TCDF	0.1	1045.95	104.5950	104.5950	333.59	33.3590	33.3590	33.3590	1024.03	102.4030								
12378-PeCDF	0.05	665.06	33.2330	33.2530	239.80	11.9900	11.9900	11.9900	673.81	33.6905								
23478-PeCDF	0.5	362.66	181.3300	181.3300	141.93	70.9650	70.9650	70.9650	356.56	178.2800								
123478-HxCDF	0.1	392.42	39.2420	39.2420	127.71	12.7710	12.7710	12.7710	243.52	24.3520								
123678-HxCDF	0.1	84.80	8.4800	8.4800	34.49	3.4490	3.4490	3.4490	54.00	5.4000								
234678-HxCDF	0.1	41.46	4.1460	4.1460	2.69	0.2690	0.2690	0.2690	20.47	2.0470								
123789-HxCDF	0.1	3.67	0.3670	0.3670	2.10	0.2100	0.2100	0.2100	3.181	3.1810								
1234678-HpCDF	0.01	601.20	6.0120	6.0120	364.28	3.6428	3.6428	3.6428	211.18	2.1118								
1234789-HpCDF	0.01	33.21	0.3321	0.3321	25.62	0.2562	0.2562	0.2562	26.44	0.2644								
12346789-OCDF	0.001	389.61	0.3896	0.3896	291.78	0.2918	0.2918	0.2918	401.07	0.4011								
nondetects = detection limit		TEQ=		TEQ=		TEQ=		TEQ=		TEQ=								
nondetects = I/2 d.l.		teq=		404.27		149.99		149.99		364.06								
nondetects = zero		teq=		404.27		149.29		149.29		363.88								
										363.69								
Moisture content (%)																		
Organic carbon (%)																		
Total organic matter (%)																		
Sample I.D.																		
C-7		C-8		C-9		C-9		C-9		C-9								
tetra-PCDF (TCDF)	1046				334						1024							
pentra-PCDF (PeCDF)	1028				382						1030							
hexa-PCDF (HxCDF)	522				167						350							
hepta-PCDF (HpCDF)	634				390						238							
octa-PCDF (OCDF)	390				292						401							
tetra-PCDD (TCDD)	4.7				ND	1.4					3							
pentra-PCDD (PeCDD)	4.3				2.2						ND							
hexa-PCDD (HxCDD)	137				69.9						51.6							
hepta-PCDD (HpCDD)	272				234						186							
octa-PCDD (OCDD)	2944				1645						1541							

Summary concentrations of the homolog g)

Concentrations of all compound (except of

Recalculation for internal standards account

* OCDF concentrations were calculated ba-

if required, OCDF concentrations can be n

ND = non detected, the detection limits wi

TABLE 9

Part 2

Part 2 (continued)

Analyte	TEF	Part 2											
		C-10				C-11				C-12			
		sampled pg/g	toxic eq. pg/g	nondetect	zero	sampled pg/g	toxic eq. pg/g	nondetect	zero	sampled pg/g	toxic eq. pg/g	nondetect	zero
2378-TCDD	1	8.00	8.0000	8.0000	ND	1.00	1.0000	0.5000	0.0000	ND	1.00	1.000	0.500
12378-PeCDD	0.5	1.50	0.7500	0.7500	0.59	0.2950	0.2950	0.2950	0.2950	ND	1.00	0.500	0.250
123478-HxCDD	0.1	82.69	8.2690	8.2690	41.84	4.1840	4.1840	4.1840	4.1840	31.65	3.165	3.165	3.165
123678-HxCDD	0.1	10.62	1.0620	1.0620	6.82	0.6820	0.6820	0.6820	0.6820	10.46	1.046	1.046	1.046
123789-HxCDD	0.1	2.82	0.2820	0.2820	ND	1.20	0.1200	0.0600	0.0000	ND	1.10	0.110	0.055
1234678-HxCDD	0.01	293.86	2.9386	2.9386	65.03	0.6503	0.6503	0.6503	0.6503	218.37	2.184	2.184	2.1837
12346789-OCDDD	0.001	4383.43	4.3834	4.3834	854.33	0.8543	0.8543	0.8543	0.8543	2337.80	2.338	2.338	2.3378
2378TCDF	0.1	1972.76	197.2760	197.2760	1376.10	137.6100	137.6100	137.6100	137.6100	725.61	72.561	72.561	72.5610
12378-PeCDF	0.05	1304.55	65.2275	65.2275	792.08	39.6040	39.6040	39.6040	39.6040	371.88	18.594	18.594	18.5940
23478-PeCDF	0.5	664.49	332.2450	332.2450	470.51	235.2550	235.2550	235.2550	235.2550	242.30	121.150	121.150	121.1500
123478-HxCDF	0.1	597.68	59.7680	59.7680	391.52	39.1520	39.1520	39.1520	39.1520	170.73	17.073	17.073	17.0730
123678-HxCDF	0.1	126.67	12.6670	12.6670	106.53	10.6530	10.6530	10.6530	10.6530	40.24	4.024	4.024	4.0240
234678-HxCDF	0.1	50.19	5.0190	5.0190	33.47	3.3470	3.3470	3.3470	3.3470	15.82	1.582	1.582	1.5820
123789-HxCDF	0.1	1.68	0.1680	0.1680	1.52	0.1520	0.1520	0.1520	0.1520	1.00	0.100	0.100	0.050
1234678-HxCDF	0.01	670.77	6.7077	6.7077	287.08	2.8708	2.8708	2.8708	2.8708	275.84	2.758	2.758	2.7584
1234789-HxCDF	0.01	53.22	0.5322	0.5322	26.62	0.2662	0.2662	0.2662	0.2662	23.43	0.234	0.234	0.2343
12346789-OCDF	0.001	434.54	0.4345	0.4345	113.81	0.1138	0.1138	0.1138	0.1138	300.27	0.300	0.300	0.3003
nondetect = detection limit		TEQ=		705.73	TEQ=		476.81	TEQ=		476.25	TEQ=		248.72
nondetects = 1/2 d.l.		teq=		705.73	teq=		475.69	teq=		475.69	teq=		247.86
nondetects = zero		teq=		705.73	teq=			teq=			teq=		247.01
Moisture content (%)													
Organic carbon (%)		24.55		28.37		28.37		23.18		23.18		23.18	
Total organic matter (%)		0.45		0.82		0.82		0.44		0.44		0.44	
Total organic carbon (%)		0.78		1.42		1.42		0.76		0.76		0.76	
Sample I.D.													
C-10		C-11		C-11		C-11		C-12		C-12		C-12	
tetra-PCDF (TCDF)		1973		1376		1376		726		726		726	
penta-PCDF (PeCDF)		1969		1263		1263		614		614		614	
hexa-PCDF (HxCDF)		776		533		533		227		227		227	
hepta-PCDF (HpCDF)		724		314		314		299		299		299	
octa-PCDF (OCDF)		435		114		114		300		300		300	
tetra-PCDD (TCDD)		8		ND		ND		1		1		1	
penta-PCDD (PeCDD)		1.5		0.6		0.6		ND		ND		ND	
hexa-PCDD (HxCDD)		96.1		48.7		48.7		42.1		42.1		42.1	
hepta-PCDD (HpCDD)		294		65		65		28		28		28	
octa-PCDD (OCDD)		4383		854		854		2338		2338		2338	

Summary concentrations of the homolog g1
 Concentrations of all compound (except of
 Recalculation for internal standards account
 * OCDF concentrations were calculated by:
 if required, OCDF concentrations can be n
 ND = non detected, the detection limits wi

TABLE 9

Part 2

Part 2 (continued)

Analyte	TEF	Part 2												Part 2												
		C-13				C-14				C-15				C-13				C-14				C-15				
		sampled pg/g	toxic eq. pg/g	nondetect	zero	sampled pg/g	toxic eq. pg/g	nondetect	zero	sampled pg/g	toxic eq. pg/g	nondetect	zero	sampled pg/g	toxic eq. pg/g	nondetect	zero	sampled pg/g	toxic eq. pg/g	nondetect	zero	sampled pg/g	toxic eq. pg/g	nondetect	zero	
2378-TcDD	1	0.54	0.540	0.540	0.540	0.55	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	0.550	
12378-PeCDD	0.5	ND	1.00	0.500	0.250	0.000	2.29	1.145	1.145	1.145	1.145	1.145	1.145	1.145	1.145	1.145	1.145	1.145	1.145	1.145	1.145	1.145	1.145	1.145	1.145	
123478-HxCDD	0.1	21.87	2.187	2.187	2.187	32.09	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	3.209	
123678-HxCDD	0.1	3.19	0.319	0.319	0.319	15.76	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	1.576	
123789-HxCDD	ND	1.00	0.100	0.050	0.000	ND	1.00	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	0.100	
1234678-HxCDD	0.01	116.71	1.167	1.167	1.167	277.48	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	2.775	
12346789-OCDDD	0.001	1293.06	1.293	1.293	1.293	3260.86	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	3.261	
2378TCDF	0.1	279.45	27.945	27.945	27.945	637.85	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	63.785	
12378-PeCDF	0.05	175.84	8.792	8.792	8.792	313.39	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670	15.670
23478-PeCDF	0.5	82.92	41.460	41.460	41.460	172.21	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	86.105	
123478-HxCDF	0.1	94.83	9.483	9.483	9.483	148.70	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	14.870	
123678-HxCDF	0.1	17.40	1.740	1.740	1.740	28.18	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	2.818	
234678-HxCDF	0.1	7.58	0.758	0.758	0.758	13.99	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	1.399	
123789-HxCDF	0.1	0.90	0.090	0.090	0.090	1.99	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	0.199	
1234678-HxCDF	0.01	156.34	1.563	1.563	1.563	290.16	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	2.902	
1234789-HxCDF	0.01	19.87	0.199	0.199	0.199	28.79	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	0.288	
12346789-OCDF	0.001	254.60	0.255	0.255	0.255	434.54	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	0.435	
nondetects = detection limit		TEQ=		98.39		TEQ=		201.09		TEQ=		201.04		TEQ=		201.04		TEQ=		961.83		961.83		961.83		
nondetects = 1/2 d.l.		teq=		98.09		teq=		97.79		97.79		97.79														
nondetects = zero																										
Moisture content (%)		29.29		30.35		30.35		30.35		30.35		30.35		30.35		30.35		30.35		30.35		30.35		30.35		
Organic carbon (%)		0.87		1.04		1.04		1.04		1.04		1.04		1.04		1.04		1.04		1.04		1.04		1.04		
Total organic matter (%)		1.5		1.79		1.79		1.79		1.79		1.79		1.79		1.79		1.79		1.79		1.79		1.79		
Sample ID		C-13		C-14		C-14		C-15		C-15		C-15														
tetra-PCDF (TCDF)		279		638		638		638		638		638		638		638		638		638		638		638		
penta-PCDF (PeCDF)		259		486		486		486		486		486		486		486		486		486		486		486		
hexa-PCDF (HxCDF)		121		193		193		193		193		193		193		193		193		193		193		193		
hepta-PCDF (HpCDF)		176		319		319		319		319		319		319		319		319		319		319		319		
octa-PCDF (OCDF)		255		435		435		435		435		435		435		435		435		435		435		435		
tetra-PCDD (TCDD)		0.5		0.5		0.5		0.5		0.5		0.5		0.5		0.5		0.5		0.5		0.5		0.5		
penta-PCDD (PeCDD)		ND		1		2.3		2.3		2.3		2.3		2.3		2.3		2.3		2.3		2.3		2.3		
hexa-PCDD (HxCDD)		251		48		48		48		48		48		48		48		48		48		48		48		
hepta-PCDD (HpCDD)		117		277		277		277																		

TABLE 9

Part 2

Part 2 (continued)

Analyte	TEF	C-16						C-17						C-18					
		sampled pg/g	toxic eq. pg/g	nondetect I/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect I/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect I/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect I/2 d.l.	nondetect zero		
2378-TCDD	1	ND	1.00	0.5000	0.0000	ND	1.00	1.000	0.5000	0.0000	ND	1.00	1.000	0.5000	0.0000	0.5000	0.0000		
12378-PeCDD	0.5	3.24	1.6200	1.6200	1.6200	ND	1.28	0.640	0.6400	0.6400	ND	1.00	0.500	0.2500	0.0000	0.2500	0.0000		
123478-HxCDD	0.1	2.42	0.2420	0.2420	0.2420	ND	1.50	0.150	0.0750	0.0750	ND	1.30	0.130	0.0650	0.0000	0.0650	0.0000		
123678-HxCDD	0.1	14.60	1.4600	1.4600	1.4600	ND	1.50	0.150	0.0750	0.0750	ND	1.30	0.130	0.0650	0.0000	0.0650	0.0000		
123789-HxCDD	0.1	3.43	0.3430	0.3430	0.3430	ND	1.49	0.149	0.1490	0.1490	ND	1.30	0.130	0.0650	0.0000	0.0650	0.0000		
1234678-HxCDD	0.01	499.92	4.9992	4.9992	4.9992	ND	9.76	0.998	0.0976	0.0976	ND	8.31	0.883	0.0831	0.0831	0.0831	0.0831		
12346789-OCDDD	0.001	7068.38	7.0684	7.0684	7.0684	ND	104.60	0.105	0.1046	0.1046	ND	83.25	0.083	0.0833	0.0833	0.0833	0.0833		
2378TCDF	0.1	27.98	2.7980	2.7980	2.7980	ND	4.38	0.438	0.4380	0.4380	ND	3.11	0.311	0.3110	0.3110	0.3110	0.3110		
12378-PeCDF	0.05	13.07	0.6535	0.6535	0.6535	ND	3.39	0.170	0.1695	0.1695	ND	1.70	0.085	0.0425	0.0000	0.0425	0.0000		
23478-PeCDF	0.5	5.82	2.9100	2.9100	2.9100	ND	1.14	0.570	0.5700	0.5700	ND	1.70	0.850	0.4250	0.0000	0.4250	0.0000		
123478-HxCDF	0.1	22.98	2.2980	2.2980	2.2980	ND	1.41	0.141	0.1410	0.1410	ND	1.00	0.100	0.0500	0.0000	0.0500	0.0000		
123678-HxCDF	0.1	5.54	0.5540	0.5540	0.5540	ND	1.30	0.130	0.0650	0.0650	ND	13.20	1.320	1.3200	1.3200	1.3200	1.3200		
234678-HxCDF	0.1	ND	0.1300	0.1300	0.1300	ND	1.00	0.100	0.0500	0.0500	ND	1.00	0.100	0.0500	0.0000	0.0500	0.0000		
123789-HxCDF	0.1	ND	1.00	0.1000	0.1000	ND	1.00	0.100	0.0500	0.0500	ND	1.00	0.100	0.0500	0.0000	0.0500	0.0000		
1234678-HxCDF	0.01	263.91	2.6391	2.6391	2.6391	ND	9.16	0.092	0.0916	0.0916	ND	5.93	0.059	0.0533	0.0533	0.0533	0.0533		
1234789-HxCDF	0.01	18.78	0.1878	0.1878	0.1878	ND	1.95	0.020	0.0098	0.0098	ND	2.65	0.027	0.0133	0.0000	0.0133	0.0000		
12346789-OCDF	0.001	675.27	0.6753	0.6753	0.6753	ND	7.55	0.008	0.0076	0.0076	ND	4.93	0.005	0.0049	0.0049	0.0049	0.0049		
nondetects = detection limit		TEQ=		29.68		TEQ=		4.06		TEQ=		5.01							
nondetects = I/2 d.l.		teq=		29.06		teq=		3.23		teq=		3.44							
nondetects = zero		teq=		28.45		teq=		2.41		teq=		1.86							
Moisture content (%)		30.26		28.54		28.54		28.54		30.65									
Organic carbon (%)		0.53		0.61		0.61		0.61		0.95									
Total organic matter (%)		1.00		1.05		1.05		1.05		1.65									
Sample I.D.		C-16		C-17		C-17		C-17		C-18									
tetra-PCDF (TCDF)		28		4.4		4.4		4.4		3.1									
penta-PCDF (PeCDF)		18.9		4.5		4.5		4.5		0									
hexa-PCDF (HxCDF)		28.5		1.4		1.4		1.4		13.2									
hepta-PCDF (HpCDF)		283		9.2		9.2		9.2		5.9									
octa-PCDF (OCDF)		675		7.5		7.5		7.5		4.9									
tetra-PCDD (TCDD)		ND		ND		ND		ND		ND		1							
penta-PCDD (PeCDD)		32		1.3		1.3		1.3		ND		1							
hexa-PCDD (HxCDD)		20.4		1.5		1.5		1.5		ND		1.3							
hepta-PCDD (HpCDD)		500		9.8		9.8		9.8		83		83							
octa-PCDD (OCDD)		7063		104.6		104.6		104.6		83.3									

Summary concentrations of the homologs g1
 Concentrations of all compound (except of
 Recalculation for internal standards account
 * OCDF concentrations were calculated based
 if required, OCDF concentrations can be non
 ND = non detected, the detection limits will

TABLE 9

Part 2

Part 2 (continued)

Analyte	TEF	C-19		13		14	
		sampled pg/g	toxic eq. pg/g	nondetect	nondetect	sampled pg/g	toxic eq. pg/g
2378-TCDD	1	ND	1.00	0.5000	0.0000	3.55	3.5500
12378-PeCDD	0.5	ND	1.00	0.500	0.2500	1.03	0.5150
123478-HxCDD	0.1	ND	1.00	0.0500	0.0500	50.84	5.0840
123678-HxCDD	0.1	ND	2.06	0.2060	0.2060	26.61	2.6610
123789-HxCDD	0.1	ND	1.15	0.115	0.0575	ND	0.140
1234678-HxCDD	0.01	ND	15.98	0.160	0.1598	1269.61	12.6961
12346789-OCDDD	0.001	ND	121.31	0.121	0.1213	18577.63	18.5778
2378TCDF	0.1	ND	36.21	3.621	0.6210	3024.20	302.4200
12378-PeCDF	0.05	ND	31.34	1.567	1.5670	1958.51	97.9255
23478-PeCDF	0.5	ND	14.61	7.305	7.3050	1156.55	578.2755
123478-HxCDF	0.1	ND	20.08	2.0080	2.0080	815.44	81.5440
123678-HxCDF	0.1	ND	1.35	0.135	0.0675	213.41	21.3410
234678-HxCDF	0.1	ND	1.35	0.135	0.0675	88.29	8.8290
123789-HxCDF	0.1	ND	1.00	0.100	0.0500	ND	1.36
1234678-HxCDF	0.01	ND	12.22	0.122	0.1222	909.54	9.0954
1234789-HxCDF	0.01	ND	2.15	0.022	0.0108	81.57	0.8157
12346789-OCDF	0.001	ND	9.02	0.009	0.0090	2288.67	2.2887
nondetect = detection limit		TEQ=		TEQ=		TEQ=	
nondetects = 1/2 d.l.		teq=		teq=		teq=	
nondetects = zero		teq=		teq=		teq=	
Moisture content (%)		29.12		27.54		24.33	
Organic carbon (%)		0.53		0.94		1.17	
Total organic matter (%)		1.00		1.63		2.01	
Sample I.D.		C-19		13		14	
tetra-PCDF (TCDF)							
penta-PCDF (PeCDF)		36.2		3024		2839	
hexa-PCDF (HxCDF)		45.9		3115		2224	
hepta-PCDF (HpCDF)		20.1		1117		823	
octa-PCDF (OCDF)		12.2		991		575	
9				2289		315	
tetra-PCDD (TCDD)		ND	1	3.6		7	
penta-PCDD (PeCDD)		ND	1	1		ND	1
hexa-PCDD (HxCDD)		2.1		77.4		77.4	
hepta-PCDD (HpCDD)		16		1270		211	
octa-PCDD (OCDD)		121		18578		1910	

Summary concentrations of the homologs
Concentrations of all compound (except of
Recalculation for internal standards account
* OCDF concentrations were calculated based
if required, OCDF concentrations can be n
ND = non detected, the detection limits will

TABLE 9

Part 2

Part 2 (continued)										Part 2									
Analyte	TEF	sampled		toxic eq.		nondetect		sampled		toxic eq.		nondetect		sampled		toxic eq.		nondetect	
		pg/g	pg/g	pg/g	pg/g	I/2 d.l.	zero	pg/g	pg/g	I/2 d.l.	zero	pg/g	pg/g	I/2 d.l.	zero	pg/g	pg/g	I/2 d.l.	zero
2378-TCDD	1	2.33	2.330	2.3300	2.3300	2.53	2.530	2.5300	2.5300	2.53	2.530	15.54	15.540	15.5400	15.5400	15.54	15.540	15.5400	15.5400
12378-PeCDD	0.5	2.10	1.050	1.0500	1.0500	ND	1.00	0.500	0.2500	0.0000	12.38	6.190	6.1900	6.1900	12.38	6.190	6.1900	6.1900	
123478-HxCDD	0.1	51.09	5.1090	5.1090	5.1090	41.24	4.124	4.1240	4.1240	41.24	4.124	12.86	12.186	12.1860	12.1860	12.86	12.186	12.1860	12.1860
123678-HxCDD	0.1	28.82	2.8820	2.8820	2.8820	3.77	0.377	0.3770	0.3770	3.77	0.377	52.62	5.262	5.2620	5.2620	52.62	5.262	5.2620	5.2620
123789-HxCDD	0.1	1.85	0.1850	0.1850	0.1850	1.99	0.199	0.1990	0.1990	1.99	0.199	15.85	1.585	1.5850	1.5850	15.85	1.585	1.5850	1.5850
1234678-HxCDD	0.01	480.00	4.8000	4.8000	4.8000	142.76	1.4276	1.4276	1.4276	142.76	1.4276	882.86	8.8286	8.8286	8.8286	882.86	8.8286	8.8286	8.8286
12346789-OCDDD	0.001	3460.10	3.4601	3.4601	3.4601	1306.73	1.3067	1.3067	1.3067	1306.73	1.3067	9539.41	9.539	9.5394	9.5394	9539.41	9.539	9.5394	9.5394
2378TCDF	0.1	2117.93	211.7930	211.7930	211.7930	5673.44	567.344	567.3440	567.3440	5673.44	567.344	691.92	69.192	69.1920	69.1920	691.92	69.192	69.1920	69.1920
12378-PeCDF	0.05	1531.82	76.5910	76.5910	76.5910	2131.69	106.585	106.585	106.585	2131.69	106.585	538.39	26.920	26.9195	26.9195	538.39	26.920	26.9195	26.9195
23478-PeCDF	0.5	886.75	443.3750	443.3750	443.3750	1413.24	706.6200	706.6200	706.6200	1413.24	706.6200	329.69	164.845	164.8450	164.8450	329.69	164.845	164.8450	164.8450
123478-HxCDF	0.1	950.83	95.0830	95.0830	95.0830	1000.74	100.074	100.074	100.074	1000.74	100.074	279.69	27.969	27.9690	27.9690	279.69	27.969	27.9690	27.9690
123678-HxCDF	0.1	209.74	20.9740	20.9740	20.9740	204.43	20.443	20.443	20.443	204.43	20.443	48.34	4.834	4.8340	4.8340	48.34	4.834	4.8340	4.8340
234678-HxCDF	0.1	85.80	8.5800	8.5800	8.5800	100.29	10.029	10.029	10.029	100.29	10.029	30.70	3.070	3.0700	3.0700	30.70	3.070	3.0700	3.0700
123789-HxCDF	0.1	1.70	0.1700	0.1700	0.1700	2.03	0.203	0.203	0.203	2.03	0.203	ND	0.0000	0.0000	0.0000	1.95	0.195	0.195	0.0075
1234678-HxCDF	0.01	554.31	5.5431	5.5431	5.5431	485.14	4.8514	4.8514	4.8514	485.14	4.8514	1236.97	12.3697	12.3697	12.3697	1236.97	12.3697	12.3697	12.3697
1234789-HxCDF	0.01	60.62	0.6062	0.6062	0.6062	61.23	0.6123	0.6123	0.6123	61.23	0.6123	77.28	0.7728	0.7728	0.7728	77.28	0.7728	0.7728	0.7728
12346789-OCDF	0.001	578.04	0.5780	0.5780	0.5780	344.26	0.3443	0.3443	0.3443	344.26	0.3443	863.59	0.864	0.8636	0.8636	863.59	0.864	0.8636	0.8636
nondetect = detection limit		TEQ=		883.11		TEQ=		1527.57		TEQ=		370.16							
nondetect = I/2 d.l.		teq=		883.11		teq=		1527.32		teq=		370.06							
nondetect = zero		teq=		882.92		teq=		1526.87		teq=		369.97							
Moisture content (%)																			
Organic carbon (%)		23.15		0.38		27.22		0.46		59.75									
Total organic matter (%)		0.66		0.8		0.8		4.05											
Sample ID																			
		15						16				23							
tetra-PCDF (TCDF)		2118		5673.0		692													
penta-PCDF (PeCDF)		2419		3545.0		868													
hexa-PCDF (HxCDF)		1248		1307.0		359													
hepta-PCDF (HpCDF)		615		546.0		1314													
octa-PCDF (OCDF)		578		344.0		864													
tetra-PCDD (TCDD)		2.3				2.5				15.5									
penta-PCDD (PeCDD)		2.1		ND		1.0				12.4									
hexa-PCDD (HxCDD)		81.8		47.0						190									
hepta-PCDD (HpCDD)		480		143.0						883									
octa-PCDD (OCDD)		3460		1307.0		9539		9539											

Summary concentrations of the homolog **g1**
 Concentrations of all compound (except of
 Recalculation for internal standards account
 * OCDF concentrations were calculated by:
 if required, OCDF concentrations can be n
 ND = non detected, the detection limits wi

TABLE 9

Part 2

Part 2 (continued)										Part 2									
Analyte	TEF	sampled		toxic eq.		nondetect		sampled		toxic eq.		nondetect		sampled		toxic eq.		nondetect	
		pg/g	pg/g	pg/g	pg/g	I/2 d.l.	zero	pg/g	pg/g	I/2 d.l.	zero	pg/g	pg/g	I/2 d.l.	zero	pg/g	pg/g	I/2 d.l.	zero
2378-TCDD	1	5.13	5.130	5.1300	5.1300	ND	1.00	1.000	0.5000	0.0000	ND	1.00	1.000	0.5000	0.0000	0.5000	0.0000	0.5000	0.0000
12378-PeCDD	0.5	ND	1.00	0.500	0.2500	0.0000	ND	1.00	0.500	0.2500	0.0000	ND	1.00	0.500	0.2500	0.0000	0.5000	0.0000	
123478-HxCDD	0.1	90.60	9.060	9.0600	9.0600	13.22	1.322	1.3220	1.3220	1.0010	1.0010	1.0010	1.0010	8.46	0.846	0.8460	0.8460	0.8460	0.8460
123678-HxCDD	0.1	18.20	1.820	1.8200	1.8200	2.71	0.2710	0.2710	0.2710	1.0350	1.0350	1.0350	1.0350	2.16	0.216	0.2160	0.2160	0.2160	0.2160
123789-HxCDD	0.1	2.71	0.271	0.2710	0.2710	168.61	1.6861	1.6861	1.6861	91.38	0.9138	0.9138	0.9138	1.30	0.130	0.1300	0.1300	0.1300	0.1300
1234678-HxCDD	0.01	168.61	1.6861	1.6861	1.6861	2017.35	2.017	2.0174	2.0174	563.92	0.5639	0.5639	0.5639	377.90	0.3778	0.3779	0.3779	0.3779	0.3779
12378TCDF	0.1	2327.82	232.782	232.7820	232.7820	232.7820	232.7820	232.7820	232.7820	559.37	55.9370	55.9370	55.9370	281.15	28.115	28.1150	28.1150	28.1150	28.1150
12378-PeCDF	0.05	1781.65	89.083	89.0825	89.0825	89.0825	89.0825	89.0825	89.0825	493.93	24.697	24.6965	24.6965	314.81	15.741	15.7405	15.7405	15.7405	15.7405
23478-PeCDF	0.5	1043.81	521.905	521.9050	521.9050	1046.97	104.697	104.6970	104.6970	273.21	136.6050	136.6050	136.6050	135.13	67.565	67.5650	67.5650	67.5650	67.5650
123478-HxCDF	0.1	1046.97	104.697	104.6970	104.6970	233.31	23.331	23.331	23.331	23.331	23.3310	23.3310	23.3310	1.51.64	15.164	15.1640	15.1640	15.1640	15.1640
123678-HxCDF	0.1	247.59	24.759	24.7590	24.7590	46.51	4.651	4.651	4.651	16.51	1.6510	1.6510	1.6510	29.45	2.945	2.9450	2.9450	2.9450	2.9450
234678-HxCDF	0.1	105.88	10.588	10.5880	10.5880	10.5880	ND	ND	ND	1.95	0.195	0.0975	0.0975	12.39	1.239	1.2390	1.2390	1.2390	1.2390
123789-HxCDF	0.1	ND	1.72	0.172	0.0860	0.0000	ND	ND	ND	121.12	1.2112	1.2112	1.2112	87.65	0.8765	0.8765	0.8765	0.8765	0.8765
1234678-HxCDF	0.01	581.90	5.819	5.8190	5.8190	163.43	1.634	1.6343	1.6343	23.47	0.2347	0.2347	0.2347	11.83	0.118	0.1183	0.1183	0.1183	0.1183
1234789-HxCDF	0.01	188.70	0.189	0.1887	0.1887	71.01	0.071	0.071	0.071	71.01	0.0710	0.0710	0.0710	42.99	0.043	0.0430	0.0430	0.0430	0.0430
nondetect = detection limit		TEQ=		1012.11		TEQ=		254.92		TEQ=		254.07		TEQ=		135.45			
nondetect = I/2 d.l.		teq=		1011.78		teq=		1011.44		teq=		253.22		teq=		134.65			
nondetect = zero														teq=		133.85			
Moisture content (%)		35.03		23.82		23.82		23.82		23.82		23.82		20.63					
Organic carbon (%)		1.63		0.29		0.29		0.51		0.51		0.51		0.11					
Total organic matter (%)		2.81												0.19					
Sample ID		37							38					39					
tetra-PCDF (TCDF)																			
pentra-PCDF (PeCDF)		2328.0															281.0		
hexa-PCDF (HxCDF)		2825.0															450.0		
hepta-PCDF (HpCDF)		1400.0															193.0		
octa-PCDF (OCDF)		745.0															99.5		
		189.0															43.0		
tetra-PCDD (TCDD)																			
pentra-PCDD (PeCDD)		5.1																	
hexa-PCDD (HxCDD)		ND	1.0														ND	1.0	
hepta-PCDD (HpCDD)		112.0																11.9	
octa-PCDD (OCDD)		169.0																47.1	
		2017.0																377.9	
Summary concentrations of the homologs g)																			
Concentrations of all compound (except of																			
Recalculation for internal standards account																			
* OCDF concentrations were calculated based on the detection limits of the internal standards.																			
If required, OCDF concentrations can be calculated based on the detection limits of the internal standards.																			
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TABLE 9
Part 2

Part 2

Summary concentrations of the homolog group
Concentrations of all compound (except of
Recalculation for internal standards account
* OCDF concentrations were calculated based
if required, OCDF concentrations can be in
ND = non detected, the detection limits will

TABLE 9

Part 2

Part 2 (continued)		Part 2										Part 2									
		55					59					60									
Analyte	TEF	sampled pg/g	toxic eq. pg/g	nondetect	nondetect																
2378-TCDD	1	4.34	4.340	4.3400	ND	1.00	1.000	0.5000	0.5000	0.0000	ND	1.10	1.100	0.5500	0.0000	ND	1.20	0.600	0.3000	0.0000	
12378-PeCDD	0.5	2.18	1.090	1.0900	ND	1.00	0.500	0.2500	0.2500	0.0000	ND	1.30	1.300	0.650	0.0000	ND	1.30	0.130	0.0650	0.0000	
123478-HxCDD	0.1	15.502	15.5020	15.5020	ND	1.30	0.130	0.0650	0.0650	0.0000	ND	1.30	0.130	0.0650	0.0000	ND	1.30	0.130	0.0650	0.0000	
123678-HxCDD	0.1	40.51	4.0510	4.0510	ND	1.30	0.130	0.0650	0.0650	0.0000	ND	1.30	0.130	0.0650	0.0000	ND	1.30	0.130	0.0650	0.0000	
123789-HxCDD	0.1	1.44	0.1440	0.1440	ND	1.30	0.130	0.0650	0.0650	0.0000	ND	1.74	0.174	0.1741	0.1741	ND	8.40	0.084	0.0840	0.0840	
12346789-OCDDD	0.001	531.96	5.320	5.3196	ND	17.41	0.174	0.1741	0.1741	0.0000	ND	8.40	0.084	0.0840	0.0840	ND	63.71	0.064	0.0637	0.0637	
2378TCDF	0.1	6670.02	6.6700	6.6700	ND	142.08	0.142	0.1421	0.1421	0.0000	ND	8.23	0.823	0.8230	0.8230	ND	6.13	0.307	0.3065	0.3065	
12378-PeCDF	0.05	551.805	5.51.8050	5.51.8050	ND	6.52	0.652	0.6520	0.6520	0.0000	ND	1.40	0.700	0.3500	0.0000	ND	1.78	0.890	0.8900	0.8900	
23478-PeCDF	0.5	3949.58	197.479	197.4790	ND	4.53	0.227	0.2265	0.2265	0.0000	ND	2.12	0.212	0.2120	0.0000	ND	3.75	0.375	0.3750	0.3750	
123478-HxCDF	0.1	2029.73	1014.865	1014.8650	ND	1.40	0.700	0.2120	0.2120	0.0000	ND	1.45	0.145	0.0725	0.0000	ND	1.60	0.160	0.0800	0.0000	
123678-HxCDF	0.1	2035.24	203.5240	203.5240	ND	1.40	0.700	0.2120	0.2120	0.0000	ND	1.00	0.100	0.0500	0.0000	ND	1.60	0.160	0.0800	0.0000	
1234678-HxCDF	0.1	37.646	37.6460	37.6460	ND	1.00	0.100	0.0500	0.0500	0.0000	ND	1.00	0.100	0.0500	0.0000	ND	2.00	0.200	0.1000	0.0000	
123789-HxCDF	0.1	14.805	14.8050	14.8050	ND	1.00	0.100	0.0500	0.0500	0.0000	ND	7.53	0.075	0.0753	0.0753	ND	5.08	0.051	0.0508	0.0508	
1234678-HxCDF	0.01	25.80	2.580	2.5800	ND	1.00	0.100	0.0500	0.0500	0.0000	ND	1.00	0.010	0.0050	0.0000	ND	2.50	0.025	0.0125	0.0000	
1234789-HxCDF	0.01	1359.68	13.597	13.5968	ND	1.00	0.010	0.0050	0.0050	0.0000	ND	6.10	0.006	0.0061	0.0061	ND	5.62	0.006	0.0056	0.0056	
nondetect = detection limit																					
nondetects = 1/2 d.l.																					
nondetects = zero																					
Moisture content (%)																					
Organic carbon (%)																					
Total organic matter (%)																					
Sample ID																					
		55																			
tetra-PCDF (TCDF)																					
pentra-PCDF (PeCDF)		5518																			
hexa-PCDF (HxCDF)		5979																			
hepta-PCDF (HpCDF)		2586																			
octa-PCDF (OCDF)		1499																			
		1523																			
tetra-PCDD (TCDD)																					
penta-PCDD (PeCDD)		4.3																			
hexa-PCDD (HxCDD)		2.2																			
hepta-PCDD (HpCDD)		197																			
octa-PCDD (OCDD)		532																			
		6670																			
Summary concentrations of the homolog gl																					
Concentrations of all compound (except of																					
Recalculation for internal standards account																					
* OCDF concentrations were calculated ba:																					
if required, OCDF concentrations can be n																					
ND = non detected, the detection limits wi																					

TABLE 9

Part 2

Part 2 (continued)

Analyte	TEF	Part 2											
		SS-1				SS-2				SS-3			
		sampled pg/g	toxic eq. pg/g	nondetect	nonzero	sampled pg/g	toxic eq. pg/g	nondetect	nonzero	sampled pg/g	toxic eq. pg/g	nondetect	nonzero
2378-TCDD	1	15.31	15.310	15.3100	15.3100	33.38	33.380	33.3800	33.3800	1.52	1.520	1.5200	1.5200
12378-PeCDD	0.5	11.89	5.945	5.9450	5.9450	17.97	8.985	8.9850	8.9850	ND	1.00	0.500	0.2500
123478-HxCDD	0.1	99.18	9.918	9.9180	9.9180	208.10	20.810	20.8100	20.8100	ND	1.30	0.130	0.0650
123678-HxCDF	0.1	58.69	5.869	5.8690	5.8690	105.33	10.533	10.5330	10.5330	ND	1.30	0.130	0.0650
123789-HxCDD	0.1	23.36	2.336	2.3360	2.3360	31.82	3.182	3.1820	3.1820	ND	1.30	0.130	0.0650
1234678-HxCDD	0.01	1016.20	10.162	10.1620	10.1620	2735.37	27.354	27.3537	27.3537	ND	36.32	0.363	0.3632
12346789-OCDDD	0.001	11122.56	11.122	11.1226	11.1226	26560.18	26.560	26.5602	26.5602	ND	369.60	0.370	0.3696
2378TCDF	0.1	950.14	95.014	95.0140	95.0140	2443.05	244.305	244.3050	244.3050	ND	13.64	1.364	1.3640
12378-PeCDF	0.05	584.12	29.206	29.2060	29.2060	1430.97	71.549	71.5485	71.5485	ND	9.07	0.454	0.4535
23478-PeCDF	0.5	356.00	178.000	178.0000	178.0000	888.76	444.380	444.3800	444.3800	ND	6.94	3.470	3.4700
123478-HxCDF	0.1	482.45	48.245	48.2450	48.2450	1059.99	105.999	105.9990	105.9990	ND	6.06	0.606	0.6060
123678-HxCDF	0.1	73.48	7.348	7.3480	7.3480	186.34	18.634	18.6340	18.6340	ND	1.29	0.129	0.1290
234678-HxCDF	0.1	51.16	5.116	5.1160	5.1160	97.83	9.783	9.7830	9.7830	ND	1.00	0.100	0.0500
123789-HxCDF	0.1	3.95	0.395	0.3950	0.3950	2.69	0.269	0.2690	0.2690	ND	1.00	0.100	0.0500
1234678-HxCDF	0.01	1973.82	19.738	19.7382	19.7382	4060.38	40.604	40.6038	40.6038	ND	15.66	0.156	0.1566
1234789-HxCDF	0.01	76.70	0.767	0.7670	0.7670	204.74	2.047	2.0474	2.0474	ND	1.90	0.019	0.0095
12346789-OCDF	0.001	2066.98	2.067	2.0670	2.0670	6795.69	6.796	6.7957	6.7957	ND	11.90	0.012	0.0119
nondetects = detection limit													
nondetects = 1/2 d.l.													
nondetects = zero													
Moisture content (%)													
Organic carbon (%)													
Total organic matter (%)													
Sample I.D.													
SS-1													
tetra-PCDF (TCDF)		950				2443						13.6	
penta-PCDF (PeCDF)		940				2320						16	
hexa-PCDF (HxCDF)		611				1347						7.3	
hepta-PCDF (HpCDF)		2051				4265						15.7	
octa-PCDF (OCDF)		2067				6796						11.9	
tetra-PCDD (TCDD)		15.3				33.4						1.5	
pentra-PCDD (PeCDD)		11.9				18						ND	1.0
hexa-PCDD (HxCDD)		181				345						ND	1.3
hepta-PCDD (HpCDD)		1016				2735						36.3	
octa-PCDD (OCDD)		11123				26560						369.6	

Summary concentrations of the homologs g1
 Concentrations of all compound (except of
 Recalculation for internal standards account
 * OCDF concentrations were calculated by:
 if required, OCDF concentrations can be n
 ND = non detected, the detection limits will

TABLE 9

Part 2

Part 2 (continued)

Analyte	TEF	SS-4		SS-5		SS-6	
		sampled pg/g	toxic eq. pg/g	nondetect I/2 d.l.	sampled pg/g	toxic eq. pg/g	nondetect I/2 d.l.
2378-TCDD	1	1.55	1.5500	1.5500	40.99	40.9900	40.9900
12378-PeCDD	0.5	ND	0.500	0.2500	27.75	13.8750	13.8750
123478-HxCDD	0.1	ND	0.100	0.0500	287.88	28.7880	28.7880
123678-HxCDF	0.1	ND	2.09	0.2090	142.84	14.2840	14.2840
123789-HxCDD	0.1	ND	1.50	0.0750	53.02	5.3020	5.3020
1234678-HxCDD	0.01	ND	61.29	0.6129	2998.75	29.9875	29.9875
12346789-OCDDD	0.001	ND	380.13	0.3801	34835.28	34.8353	34.8353
2378TCDF	0.1	1.52	0.1520	0.1520	1315.76	131.5760	131.5760
12378-PeCDF	0.05	ND	1.44	0.0720	699.72	34.9860	34.9860
23478-PeCDF	0.5	ND	1.00	0.5000	456.51	228.2550	228.2550
123478-HxCDF	0.1	ND	7.49	0.7490	619.14	61.9140	61.9140
123678-HxCDF	0.1	ND	1.00	0.0500	115.82	11.5820	11.5820
234678-HxCDF	0.1	ND	1.00	0.0500	68.36	6.8360	6.8360
123789-HxCDF	0.1	ND	1.00	0.0500	1.70	0.1700	0.1700
1234678-HxCDF	0.01	ND	38.05	0.3805	4131.74	41.3174	41.3174
1234789-HxCDF	0.001	ND	2.25	0.023	0.0113	0.0006	0.0006
12346789-OCDF	0.001	ND	18.83	0.019	0.0188	0.0188	0.0188
nondetects = detection limit							
nondetects = I/2 d.l.							
nondetects = zero							
Moisture content (%)							
Organic carbon (%)							
Total organic matter (%)							
Sample ID							
		SS-4			SS-5		SS-6
tetra-PCDF (TCDF)							
penta-PCDF (PeCDF)		1.5			1316		832
hexa-PCDF (HxCDF)		1.4			1156		724
hepta-PCDF (HpCDF)		7.5			805		289
octa-PCDF (OCDF)		38.0			4375		320
octa-PCDF (OCDD)		18.8			7565		227
tetra-PCDD (TCDD)							
penta-PCDD (PeCDD)		ND	1.0		41		2.2
hexa-PCDD (HxCDD)		2.1			27.8		1.2
hepta-PCDD (HpCDD)		61.3			484		38.3
octa-PCDD (OCDD)		380.1			2999		221
					34835		2462

Summary concentrations of the homolog groups
 Concentrations of all compound (except of
 Recalculation for internal standards account
 * OCDF concentrations were calculated based on
 if required, OCDF concentrations can be non-detect limits will be
 ND = non detected, the detection limits will be

TABLE 9

Part 2

Part 2 (continued)

Analyte	TEF	Part 2										Part 2											
		SS-7					SS-7r					SS-8					SS-8						
		sampled	toxic eq.	nondetect	sampled	toxic eq.	nondetect	sampled	toxic eq.	nondetect	sampled	toxic eq.	nondetect	sampled	toxic eq.	nondetect	sampled	toxic eq.	nondetect	sampled	toxic eq.		
		pg/g	pg/g	I/2 d.l.	zero	pg/g	pg/g	I/2 d.l.	zero	pg/g	pg/g	I/2 d.l.	zero	pg/g	pg/g	I/2 d.l.	zero	pg/g	pg/g	I/2 d.l.	zero		
2378-TCDD	1	5.71	5.710	5.7100	5.7100	7.80	7.800	7.8000	7.8000	4.25	4.250	4.2500	4.2500	3.71	1.855	1.8550	1.8550	3.71	1.855	1.8550	1.8550		
12378-PeCDD	0.5	5.08	2.540	2.5400	2.5400	4.17	2.085	2.0850	2.0850	5.130	5.130	5.1300	5.1300	19.15	1.915	1.9150	1.9150	4.93	0.493	0.4930	0.4930		
123478-HxCDD	0.1	7.78	7.778	7.7780	7.7780	86.94	8.694	8.6940	8.6940	6.8480	6.8480	6.8480	6.8480	20.32	2.032	2.0320	2.0320	422.36	4.224	4.2236	4.2236		
123678-HxCDD	0.1	85.83	8.583	8.5830	8.5830	68.48	6.848	6.8480	6.8480	2.0320	2.0320	2.0320	2.0320	917.28	9.173	9.1728	9.1728	12.090	1.090	1.0900	1.0900		
1234678-HxCDD	0.1	39.06	3.9060	3.9060	3.9060	120.90	12.090	12.0901	12.0901	12.090	12.090	12.090	12.090	12.090	12.090	12.090	12.090	12.090	12.090	12.090	12.090		
12346789-OCDD	0.001	9453.71	9.4537	9.4537	9.4537	11577.69	11.5778	11.5778	11.5778	3274.44	327.444	327.4440	327.4440	106.80	10.680	10.6801	10.6801	106.80	10.680	10.6801	10.6801		
2378TCDF	0.1	3920.61	392.061	392.0610	392.0610	392.0610	392.0610	392.0610	392.0610	83.06	8.306	8.3060	8.3060	10.663	1.0663	1.06630	1.06630	10.663	1.0663	1.06630	1.06630		
12378-PeCDF	0.05	1917.22	95.861	95.8610	95.8610	1997.95	99.898	99.898	99.898	3.07	0.307	0.3070	0.3070	1137.82	568.910	568.9100	568.9100	1214.53	607.265	607.2650	607.2650		
23478-PeCDF	0.5	1131.51	11.3151	11.3151	11.3151	1251.47	12.515	12.515	12.515	1251.47	12.515	12.515	12.515	129.21	1.292	1.2921	1.2921	112.18	1.122	1.1218	1.1218		
123478-HxCDF	0.1	859.23	0.859	0.8592	0.8592	2231.17	2.231	2.231	2.231	2.2312	2.2312	2.2312	2.2312	25.461	2.5461	2.54610	2.54610	197.43	19.743	19.7430	19.7430		
123678-HxCDF	0.1	106.63	10.663	10.6630	10.6630	83.06	8.306	8.3060	8.3060	8.3060	8.306	8.3060	8.3060	10.663	1.0663	1.06630	1.06630	10.663	1.0663	1.06630	1.06630		
1234678-HxCDF	0.1	3.94	0.394	0.3940	0.3940	3.07	0.307	0.3070	0.3070	3.07	0.307	0.3070	0.3070	1131.51	11.3151	11.3151	11.3151	1251.47	12.515	12.515	12.515		
1234789-HxCDF	0.001	129.21	1.292	1.2921	1.2921	112.18	1.122	1.1221	1.1221	112.18	1.122	1.1218	1.1218	1.2921	0.8592	0.8592	0.8592	1.2921	1.2921	1.2921	1.2921		
nondetects = detection limit		TEQ=	1261.12	1261.12	1261.12	TEQ=	1233.84	1233.84	1233.84	TEQ=	1233.84	1233.84	1233.84	1261.12	teq=	teq=	teq=	teq=	teq=	teq=	teq=		
nondetects = I/2 d.l.		teq=				teq=			teq=														
nondetects = zero		teq=				teq=			teq=														
Moisture content (%)			24.18	3.38	5.82					24.11	3.21	5.54					24.07		1.19		2.05		
Organic carbon (%)																							
Total organic matter (%)																							
Sample I.D.			SS-7							SS-7r							SS-8						
tetra-PCDF (TCDF)																							
pentra-PCDF (PeCDF)																							
hexa-PCDF (HxCDF)																							
hepta-PCDF (HpCDF)																							
octa-PCDF (OCDF)																							
tetra-PCDD (TCDD)																							
pentra-PCDD (PeCDD)																							
hexa-PCDD (HxCDD)																							
hepta-PCDD (HpCDD)																							
octa-PCDD (OCDD)																							

Summary concentrations of the homologs
Concentrations of all compound (except of
Recalculation for internal standards account
* OCDF concentrations were calculated based
if required, OCDF concentrations can be non-detectable
ND = non detected, the detection limits will be

TABLE 9

Part 2

Part 2 (continued)				SS-9						SS-10			
Analyte	TEF	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero	sampled pg/g	toxic eq. pg/g	nondetect 1/2 d.l.	nondetect zero
23,78-TcDD	1	4.58	4.580	4.5800	4.5800	ND	1.00	1.000	0.5000	0.0000			
12,378-PeCDD	0.5	4.39	2.195	2.1950	2.1950	ND	0.72	0.360	0.3600	0.3600			
12,3478-HxCDD	0.1	71.64	7.164	7.1640	7.1640	ND	1.20	0.120	0.0600	0.0000			
12,3678-HxCDD	0.1	33.44	3.344	3.3440	3.3440		1.50	0.150	0.1500	0.1500			
12,3789-HxCDD	0.1	7.17	0.717	0.7170	0.7170		1.37	0.137	0.1370	0.1370			
12,34678-HpCDD	0.01	574.20	5.742	5.7420	5.7420		11.73	0.117	0.1173	0.1173			
12,346789-OCDD	0.001	630.70	6.321	6.3207	6.3207		103.53	0.104	0.1035	0.1035			
23,787CDF	0.1	3406.24	340.624	340.6240	340.6240		1.00	0.100	0.1000	0.1000			
12,378-PeCDF	0.05	3084.30	154.215	154.2150	154.2150		1.21	0.061	0.0605	0.0605			
23,478-PeCDF	0.5	1460.63	730.315	730.3150	730.3150		1.13	0.565	0.5650	0.5650			
12,3478-HxCDF	0.1	1749.01	174.901	174.9010	174.9010	ND	1.00	0.100	0.0500	0.0000			
12,3678-HxCDF	0.1	3118.54	311.854	311.8540	311.8540	ND	1.00	0.100	0.0500	0.0000			
23,4678-HxCDF	0.1	116.26	11.626	11.6260	11.6260	ND	1.00	0.100	0.0500	0.0000			
12,3789-HxCDF	0.1	13.30	1.330	1.3300	1.3300	ND	1.00	0.100	0.0500	0.0000			
12,34678-HpCDF	0.01	1243.62	12.436	12.4362	12.4362		12.4362	3.91	0.039	0.0391			
12,34789-HpCDF	0.01	125.09	1.251	1.2509	1.2509	ND	2.00	0.020	0.0100	0.0000			
12,346789-OCDF	0.001	1782.44	1.782	1.7824	1.7824	ND	3.40	0.003	0.0034	0.0000			
nondetects = detection limit		TEQ= 1490.40		TEQ= 1490.40		TEQ= 1490.40		TEQ= 1490.40		TEQ= 1490.40		TEQ= 1490.40	
nondetects = 1/2 d.l.		teq= teq=		teq= teq=		teq= teq=		teq= teq=		teq= teq=		teq= teq=	
nondetects = zero													
Moisture content (%)		18.58						25.38					
Organic carbon (%)		2.24						3.56					
Total organic matter (%)		3.85						6.14					
Sample ID.		SS-9				SS-10							
tetra-PCDF (TCDF)		3406				ND		1.0					
penta-PCDF (PeCDF)		4545				ND		2.3					
hexa-PCDF (HxCDF)		2197				ND		1.0					
hepta-PCDF (HpCDF)		1369				ND		3.9					
octa-PCDF (OCDF)		1782				ND		3.4					
tetra-PCDD (TCDD)		4.6				ND		1.0					
penta-PCDD (PeCDD)		4.4				ND		0.7					
hexa-PCDD (HxCDD)		112				ND		2.9					
hepta-PCDD (HpCDD)		574				ND		11.7					
octa-PCDD (OCDD)		6321				ND		104.0					

Summary concentrations of the homolog g1
 Concentrations of all compound (except of
 Recalculation for internal standards account
 * OCDF concentrations were calculated ba:
 If required, OCDF concentrations can be n
 ND = non detected, the detection limits wil

TABLE 9
Part 3

DATA FLAGS

In order to assist with data interpretation, data qualifier flags are used on the final reports.

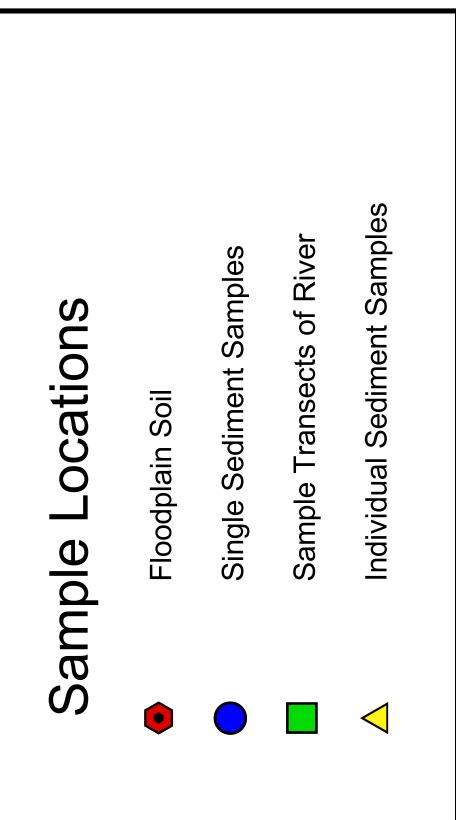
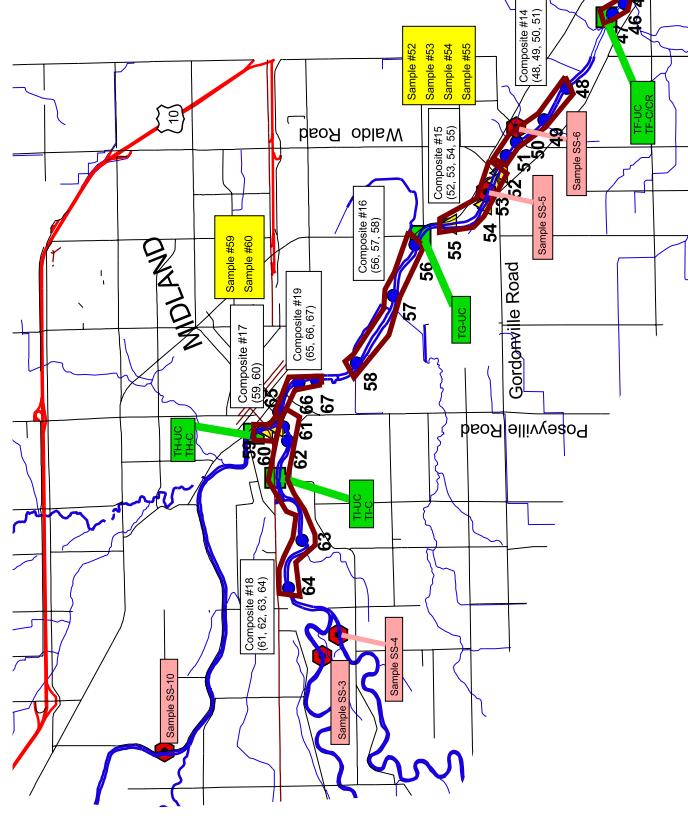
The most commonly used flags are:

- ND** = analyte not detected. Value is the detection limit.
- B** = analyte has been detected in the laboratory method blank as well as in an associated field sample.
- E** = indicates a concentration based on an analyte to internal standard ratio which exceeds the range of the calibration curve. Values which are outside the calibration curve are estimates only.
- I** = indicates labeled standards have been interfered with on the GC column by coeluting, interferent peaks.
- J** = indicates a concentration based on an analyte to internal standard ration which is below the calibration curve. Values outside the calibration curve are estimates only.
- PR** = indicates that a GC peak is poorly resolved. The concentrations or amounts reported for such peaks are most likely overestimated.
- Q** = indicates the presence of QC ion instabilities caused by quantitative interferences.
- S** = indicates that the response of a specific PCDD/PCDF isomer has exceeded the normal dynamic range of the mass spectrometer detection system. The corresponding signal is saturated and the reported analyte concentration is a 'minimum estimate'. Results for saturated analytes are reported as greater than the upper calibration limit.
- U** = indicates that a specific isomer cannot be resolved from a large, coeluting interferent GC peak. The specific isomer is reported as not detected as a valid concentration cannot be determined. The calculated detection limit, therefore, should be considered an underestimated value.
- V** = indicates that, although the percent recovery of a labeled standard may be below a specific QC limit, the signal-to-noise ratio of the peak is greater than ten-to-one. The standard is considered reliably quantifiable. All quantitations derived from the standard are considered valid as well.
- X** = indicates that a polychlorodibenzofuran (PCDF) peak has eluted at the same time as the associated diphenyl ether (DPE) and that the DPE peak intensity is at least ten percent of the total PCDF peak intensity. Total PCDF values are flagged "X" if the total DPE contribution to the total PCDF value is greater than ten percent.

FIGURES

Figure 3

Saginaw Bay Watershed Sediment Study Tittabawassee River Midland & Saginaw Counties, Michigan

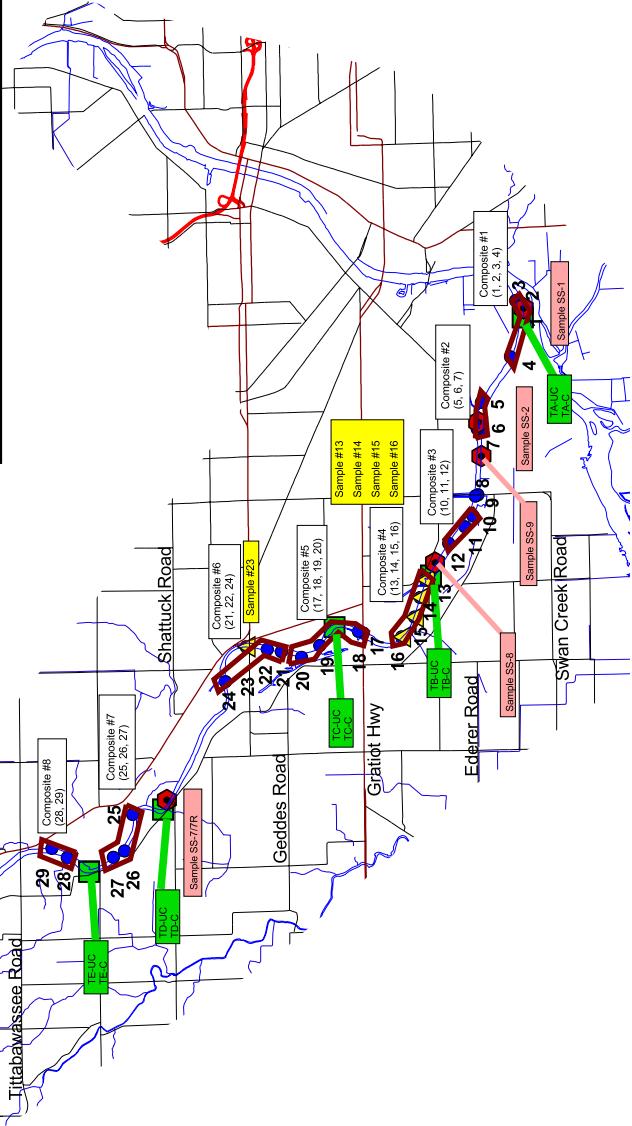


Floodplain Soil

Single Sediment Samples

Sample Transects of River

Individual Sediment Samples



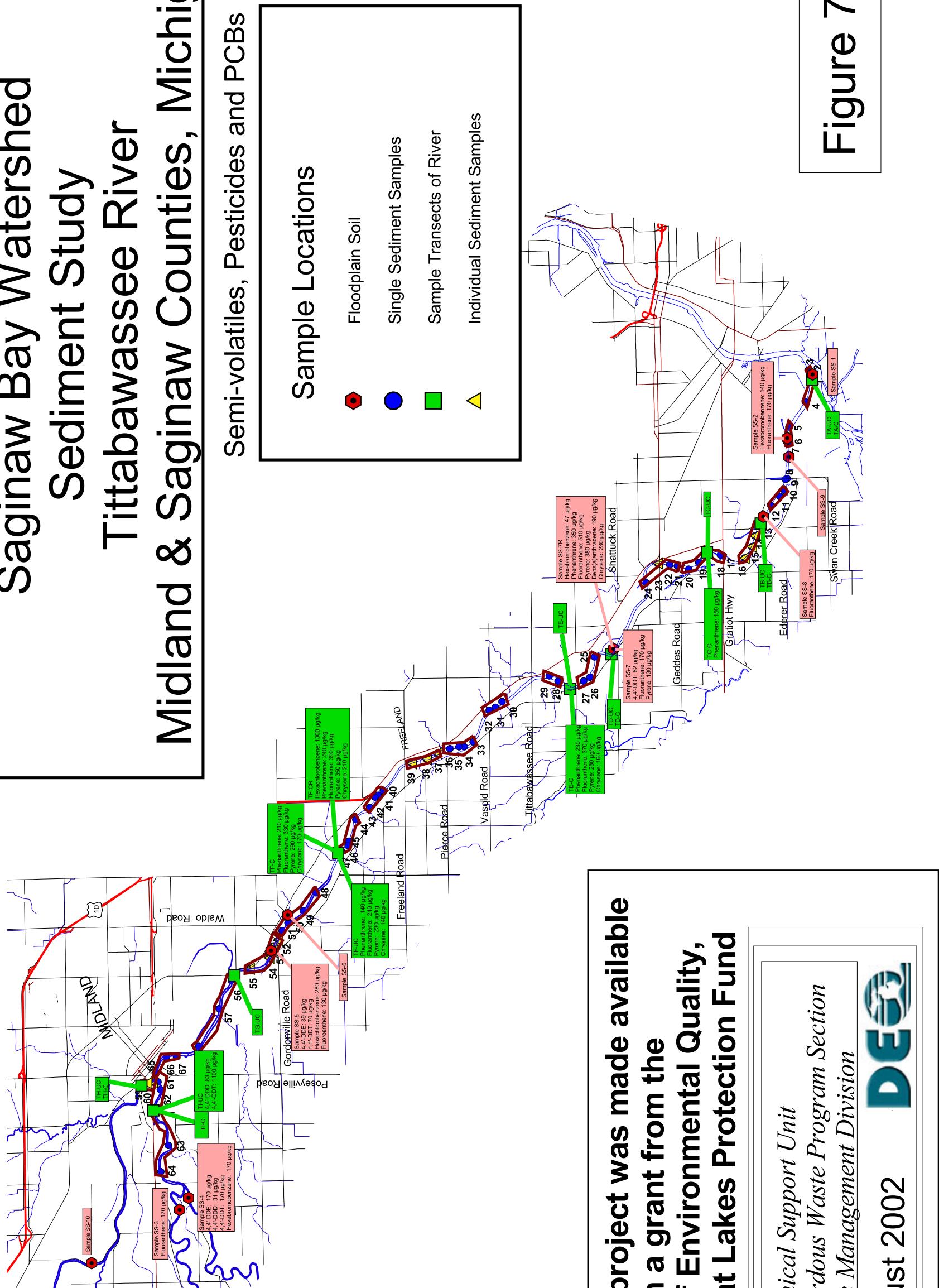
**Funding for this project was made available
through a grant from the
Department of Environmental Quality,
Michigan Great Lakes Protection Fund**

W **N**
D **E**
Technical Support Unit
Hazardous Waste Program Section
Waste Management Division

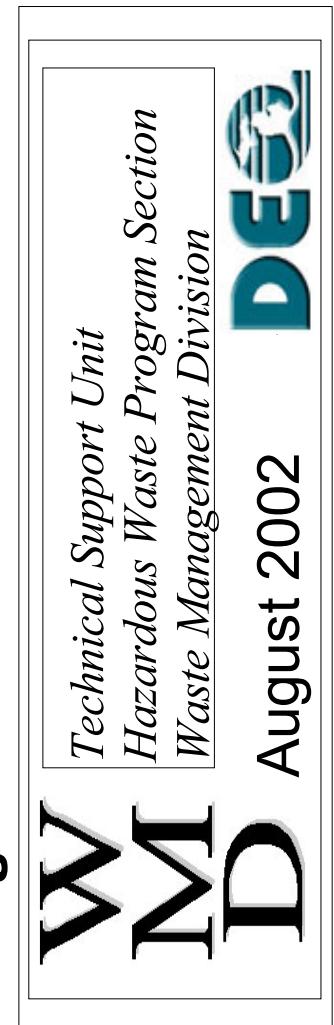


August 2002

Saginaw Bay Watershed Sediment Study Tittabawassee River Midland & Saginaw Counties, Michigan



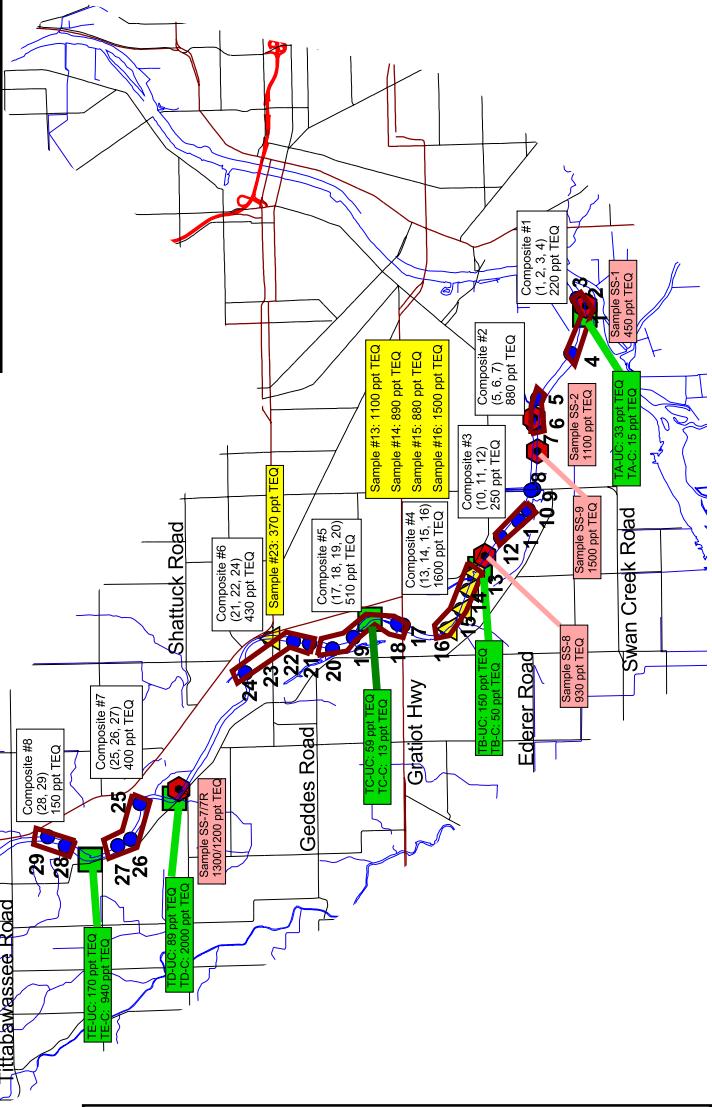
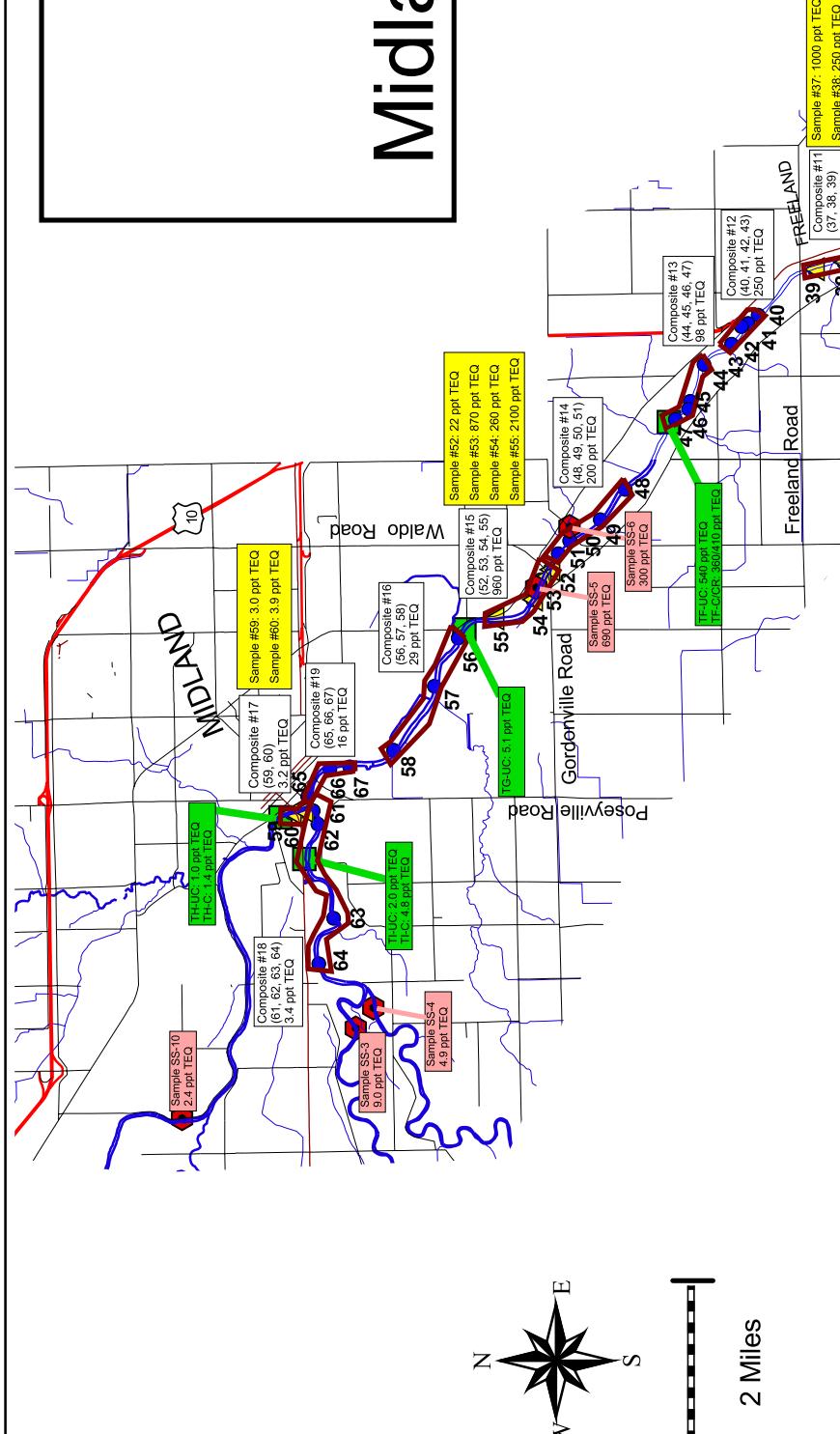
**Funding for this project was made available
through a grant from the
Department of Environmental Quality,
Michigan Great Lakes Protection Fund**



August 2002

Figure 7

Saginaw Bay Watershed Sediment Study Tittabawassee River Midland & Saginaw Counties, Michigan



**Funding for this project was made available
through a grant from the
Department of Environmental Quality,
Michigan Great Lakes Protection Fund**

W **N**
D August 2002

Technical Support Unit
Hazardous Waste Program Section
Waste Management Division

Figure 8



Figure 9

TITTABAWASSEE RIVER STUDY

Transect Analyses

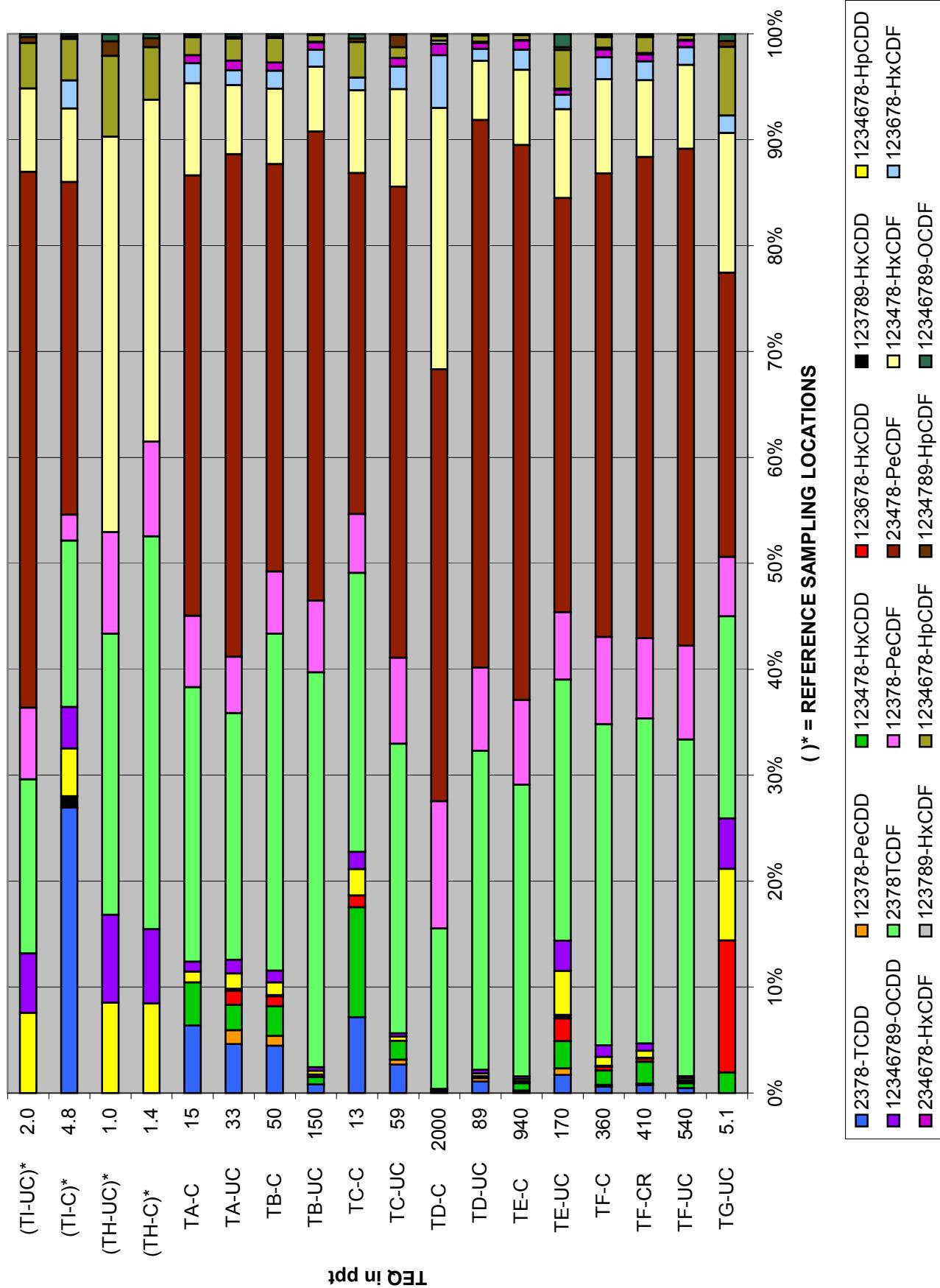


Figure 10

TITTABAWASSEE RIVER STUDY

Composite Analyses

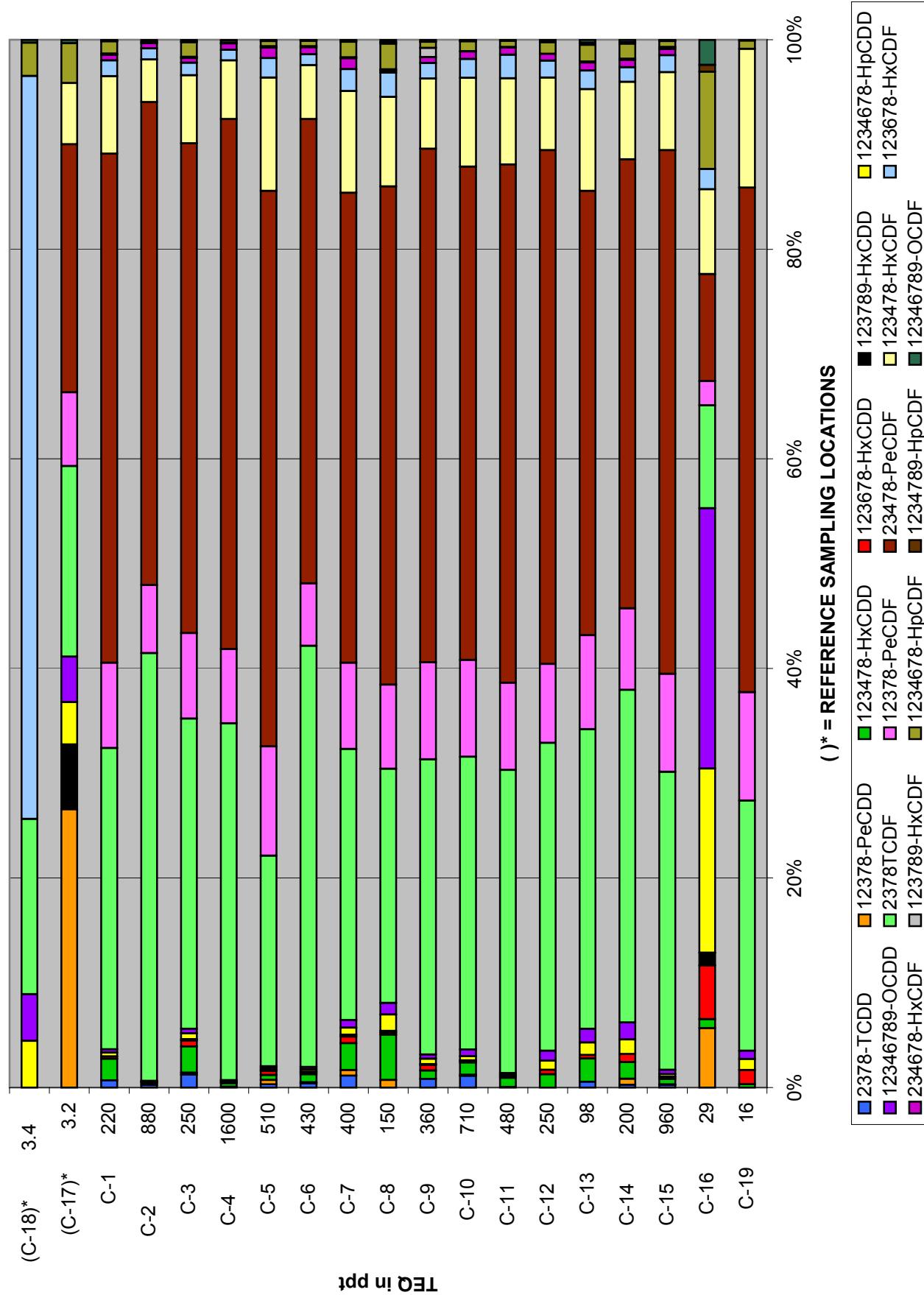


Figure 11

TITTABAWASSEE RIVER STUDY

Individual Analyses

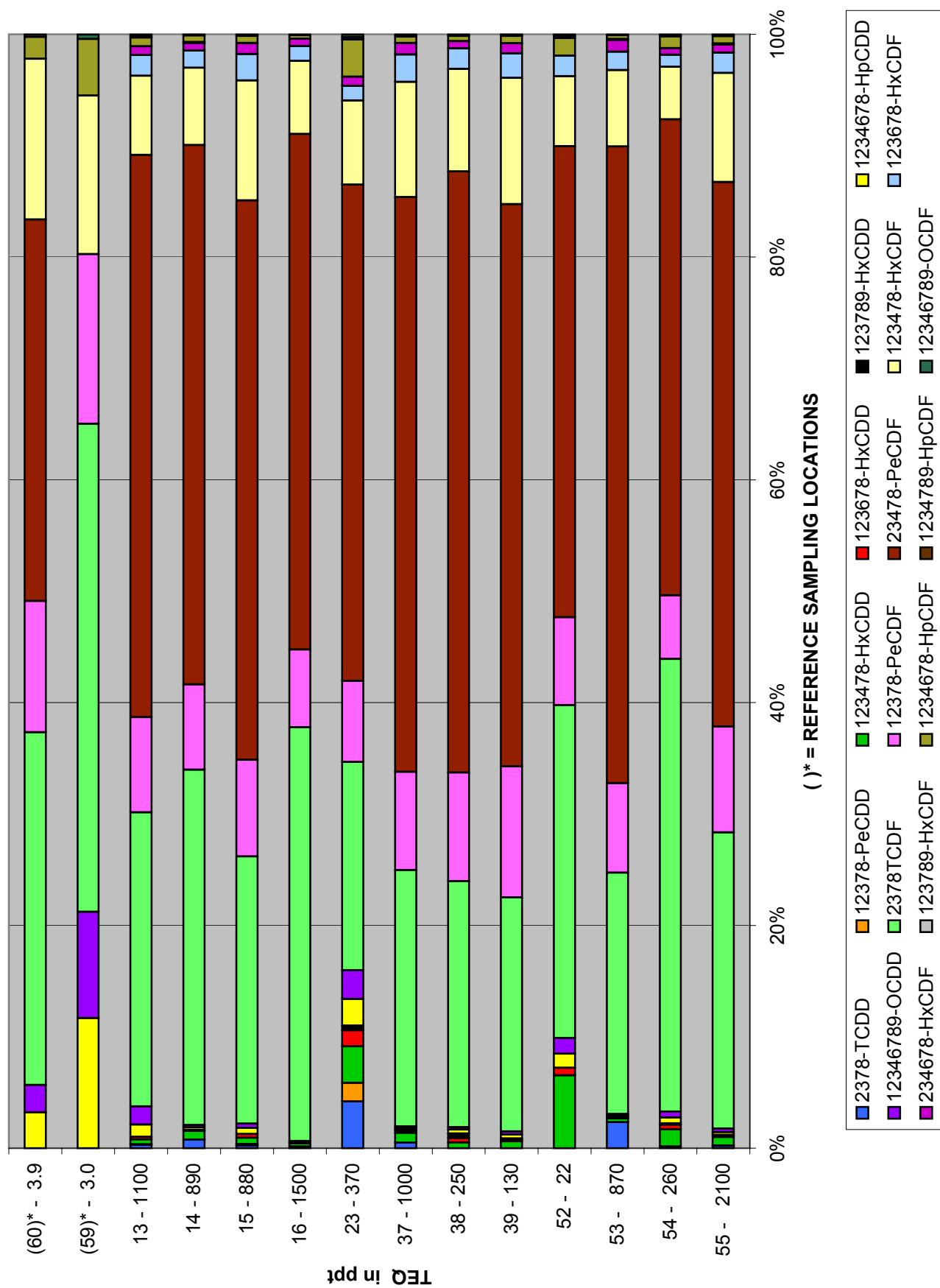
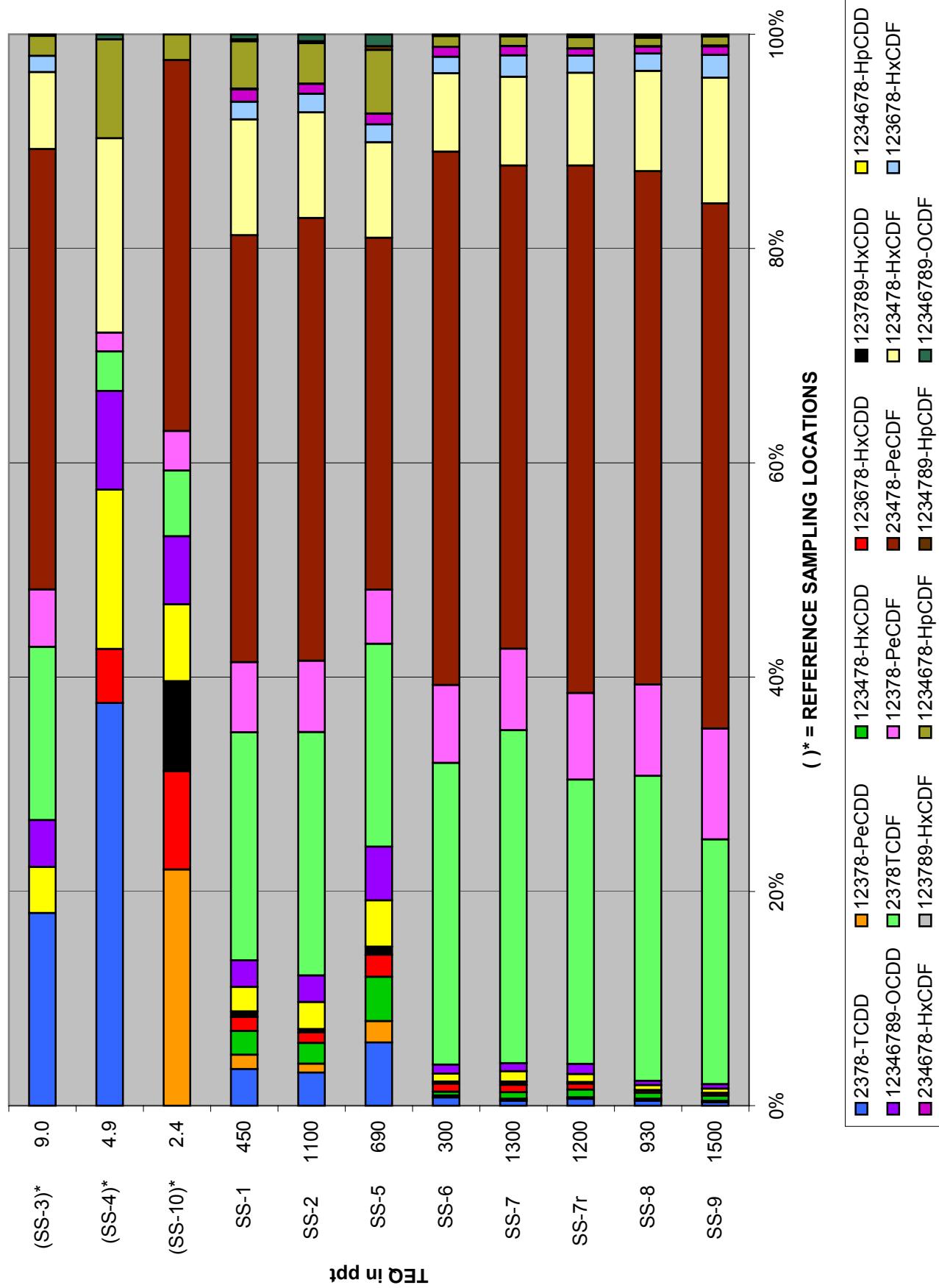


Figure 12

Flood Plain Soils

TITTABAWASSEE RIVER STUDY



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References Not Included in Appendix 2

- De Rosa, C.T.; Brown, D.; Dhara, R.; Garrett, W.; Hansen, H.; Holler, J.; Jones, D.; Jordan-Izaguirre, D.; O'Connor, R.; Pohl, H.; Xintaras, C. Dioxin and dioxin-like compounds in soil, Part 1:ATSDR interim policy guideline. *Agency for Toxic Substances and Disease Registry, U.S. Department of Health and Human Services Atlanta, Georgia* **1997**, 13, 759-768.
- Hilscherova, K., Machala, M., Kannan, K., Blankenship, A.L. and Giesy, J.P. (2000). *Environ. Sci. Pollut. Res.*, 7, 159-171.
- Hilscherova, K., Kannan, K., Kang, Y-S., Holoubek, I., Machala, M., Masunaga, S., Nakanishi, J. and Giesy, J.P. (2001). *Environ. Toxicol. Chem.* 20, 2768-2777.
- Im, S.H., Kannan, K., Matsuda, M., Giesy, J.P. and Wakimoto, T. (2002). *Environ. Toxicol. Chem.*, 21, 245-252.
- Kannan, K., Kober, J.L., Kang, Y-S., Masunaga, S., Nakanishi, J., Ostaszewski, A. and Giesy, J.P. (2001). *Environ. Toxicol. Chem.*, 20, 1878-1889.
- Kannan, K., Villeneuve, D.L., Yamashita, N., Imagawa, T., Hashimoto, S., Miyazaki, A. and Giesy, J.P. (2000). *Environ. Sci. Technol.* 34, 3560-3567.
- Kannan, K., Watanabe, I. and Giesy, J.P. (1998). *Toxicol. Environ. Chem.*, 67, 135-146.
- Khim J.S., Villeneuve D.L., Kannan K., Koh C.H., and Giesy J.P. (1999). *Environ Sci Technol* 33:4199-4205.
- Khim, J.S., Lee, K.T., Villeneuve, D.L., Kannan, K., Giesy, J.P. and Koh, C.H. (2001). *Arch. Environ. Contam. Toxicol.*, 40, 151-160.
- Masunaga, S., Takasuga, T. and Nakanishi, J. (2001). Dioxin and dioxin-like PCB impurities in some Japanese agrochemical formulations. *Chemosphere*, 44, 873-885.
- Swami, K., Narang, A.S., Narang, R.S. and Eadon, G.A. (1992). Thermally induced formation of PCDD and PCDF from tri- and tetrachlorobenzene in dielectric fluids. *Chemosphere*, 24, 1845-1853.
- Villeneuve DL, Blankenship AL, Giesy JP. 2000. Derivation and application of relative potency estimates based on in vitro bioassay results. *Environ Toxicol Chem* 19, 2835-2843.
- Villeneuve, D.L., Khim, J.S., Kannan, K., Giesy, J.P. 2000. Aquat. Toxicol., 54, 125-141.

Wakimoto, T., Kannan, N., Ono, M., Tatsukawa, R. and Masuda, Y. (1988).
Isomer-specific determination of polychlorinated dibenzofurans in Japanese
and American polychlorinated biphenyls. *Chemosphere*, 17, 743-750.

Yamashita, N., Kannan, K., Imagawa, T., Villeneuve, D.L., Hashimoto, S.,
Miyazaki, A. and Giesy, J.P. 2000. *Environ. Sci. Technol.*, 34, 3560-3567.